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Patentanmeldung Nr. Patent application No. Demande de brevet n°

03290834.5

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Anmeldung Nr:
Application no.: 03290834.5
Demande no:

Anmeldetag:
Date of filing: 03.04.03
Date de dépôt:

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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
If no title is shown please refer to the description.
Si aucun titre n'est indiqué se référer à la description.)

Therapeutic products with enhanced ability to immunomodulate cell functions

In Anspruch genommene Priorität(en) / Priority(ies) claimed /Priorité(s)
revendiquée(s)
Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/
Classification internationale des brevets:

C07K16/00

Am Anmeldetag benannte Vertragsstaaten/Contracting states designated at date of
filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HU IE IT LU MC NL
PT RO SE SI SK TR LI

THERAPEUTIC PRODUCTS WITH ENHANCED ABILITY TO IMMUNOMODULATE CELL FUNCTIONS

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The present invention relates to a method for the production and the selection of human or chimæric or humanized antibodies or molecules that comprise the Fc region of human IgG, capable of modulating the activity of one or several particular Fc receptors, such as the triggering of inhibitory functions through the human type II receptors of IgG
10 (FcgammaRII/CD32).

INTRODUCTION

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Antibody based therapy in human has emerged with the possibility of producing partially human antibodies (chimeric antibodies) and more recently humanized antibodies. Humanized or murine/human chimeric recombinant antibodies have been developed to prevent the appearance of human anti-mouse antibodies ("HAMA") in patients treated with mouse antibodies, thus avoiding side-effects due to the formation of immune-complexes between HAMA and the injected antibody. In addition, such engineered antibodies exhibit
20 stronger effector functions than their murine counterparts, as their binding to human FcgammaR is improved. Numerous antibodies are currently being tested in clinical trials for the treatment of cancer for example. But, as of today, only few antibodies have shown efficacy at low dose and several trials have been prematurely terminated. Higher dose antibody administration is mainly limited by two factors. First, it increases likewise
25 numerous side effects. Second, it is still a problem to produce large quantities of antibodies and the cost associated with scaling-up is not always economically viable.

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These problems need to be addressed before applying antibodies therapy to large population afflicted with various diseases.

It has been shown that the Fc region of IgG is essential for the functions of antibodies. The recruitment of C1q and of membrane FcγR through the Fc region of IgG allows to trigger various effector mechanisms such as cytotoxicity, cytokine release or endocytosis. For instance, it has been shown that human IgG1, the most represented human IgG subclass, that comprises IgG1, IgG2, IgG3 and IgG4, triggers the highest antibody dependent cell cytotoxicity (ADCC). It is due to the ability of human IgG1 to efficiently bind to FcγR expressed on NK cells and monocytes/macrophages. In addition, FcγR play an important role in immune regulation by triggering inhibitory functions. Three distinct classes of FcγR have been defined both in humans and mice. Human FcγR include the high affinity FcγRI (CD64), and the low-affinity FcγRII (CD32) and FcγRIII (CD16). Three genes encode FcγRI isoforms (A, B, C), three genes encode FcγRII isoforms (A1, B1, B2) and two genes encode FcγRIII isoforms (A, B). Attempts have been made to improve the efficacy of IgG antibodies by modifying the amino-acid sequence in the Fc domain so that a modulation of the interactions of Fc region with FcγR can be achieved (see for example WO 99/54572).

Biophysical and molecular studies have indicated that several amino-acid residues located in the hinge region between the CH1 and CH2 domains and immediately adjacent to the N terminus of the CH2 domain of IgG1, as well as the sugar chain linked to the CH2 domain at position Asn²⁹⁷, play a critical role in FcγR binding. On the one hand, a common set of IgG1 residues is involved in binding to all FcγR (I, II, III), but residues outside this common set were identified when FcγRII and FcγRIII interactions with human Fcγ1 were studied in details (Shields et al., J. Biol. Chem., 276, 6591-6604, 2001). On the other hand, although not in direct contact with the FcγR, the carbohydrate attached to the conserved residue Asn²⁹⁷ on Fc is likely to stabilize the conformation of the receptor binding epitope on Fc (Radaev et al., J. Biol. Chem., 276, 16469-16477, 2001). It has been hypothesized that deglycosylation causes a conformational change in the relative orientation of the two CH2 domains such as the Fc transitions from an open to a closed conformation preventing FcγR binding (Radaev & Sun, J. Biol. Chem., 276, 16478-16483, 2001).

Analysis of IgG1 glycoforms bearing consecutively truncated oligosaccharides confirmed that removal of sugar residues permits the mutual approach of CH2 domains resulting in the generation of a closed conformation (Krapp et al., J. Mol. Biol., 325, 979-989, 2003).

- 5 Engineered IgG glycoforms have been shown to trigger optimized ADCC through the recruitment of FcγRIII. First, Umana et al. proposed that an IgG1 antibody engineered to contain increasing amounts of bisected complex oligosaccharides (bisecting N-acetylglucosamine, GlcNAc) would allow to trigger a strong ADCC as compared to its parental counterpart (Umana et al., Nature Biotechnol., 17, 176-180, 1999), although this
10 claim has been challenged (Shinkawa et al., J. Biol. Chem, 2002).

Such contradictory results show that it is difficult to identify the actual optimized oligosaccharides structures responsible for the activation or inhibition of a given FcR. We found that it is more relevant to identify patterns involved in improved binding rather than
15 focusing on a unique structure. Indeed, we have observed that binding occurs within a range of similar structures.

In this regard, we have demonstrated that particular glycosylation patterns attached to the Fc domain are responsible for enhancing Fc region - FcγRIII receptors interaction. In this regard,
20 we filed WO 01/77181 showing that it is possible to prepare compositions of antibodies with improved ADCC properties from particular cell lines, such as YB2/0, and demonstrating the role of particular glycosylation patterns. Such antibodies present short oligosaccharide chains, a weak sialylation, no or weak level of bisecting GlcNAc and a low level of fucose between 20% and 50%.

25 It was also postulated in WO 00/61739 that the presence or the absence of fucose modulates the activity of antibodies. The lack of fucose on human IgG1 N-linked oligosaccharides has been shown to improve FcγRIII binding and ADCC. Recombinant human IgG1 produced in YB2/0 cells (Shinkawa et al., J. Biol. Chem., 2002) or in CHO-Lec13 cells
30 (Shields et al., J. Biol. Chem., 277, 26733-26740, 2002), which exhibited a low-fucose

content or were deficient in fucose as compared to the same IgG1 produced in wild-type CHO cells, showed an enhanced ability to trigger cellular cytotoxicity. By contrast, correlation between galactose and ADCC was not observed and the content of bisecting GlcNAc only marginally affected ADCC (Shinkawa et al., J. Biol. Chem., 2002).

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While the above results confirm what we previously described in our application WO 01/77181, we now have evidence showing that the alteration of antibody activity can not be resumed solely at the fucose level. The implication of GlcNAc, mannose, sialic acid, galactose and their respective position is extremely variable in antibodies produced from different cell lines. This observation confirms that different glycosylation patterns are not only involved in Fc region - Fc receptors interaction but are also responsible for the function of the antibody.

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Therefore, the general purpose of the invention is to provide a method for fine-tuning the function of a given antibody in respect to the different Fc receptors. Different defined patterns would allow the modulation of the different FcR.

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Among the receptors for the Fc region of IgG, type IIB Fcγ receptors is of particular interest. FcγRII are single-chain, low-affinity receptors that bind with increased affinity IgG present in immune-complexes or bound to cell surface antigens. Genetic and protein analyses have shown that FcγRII are present under two isoforms in Human (FcγRIIB1 and FcγRIIB2), generated by alternative splicing of the three exons that encode the intracytoplasmic domain of the receptor. FcγRIIB are expressed on B cells, monocytes, dendritic cells, mast cells and basophils. By contrast to all the other known FcγR, FcγRIIB isoforms inhibit cell activation induced through activating receptors (Amigorena et al., Science, 256, 1808-1812, 1992). Molecular analyses have shown that the presence of an YxxL motif, termed "immunoreceptor tyrosine-based inhibition motif" (ITIM) in the intracellular domain of FcγRIIB is directly involved in the inhibitory function of the receptor. When tyrosyl phosphorylated, ITIM binds SH domain of phosphatases, namely SHP-1, SHP-2 and the phosphatidylinositol polyphosphatase.

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5-phosphatase, SHIP. SHIP can dephosphorylate PIP3 into PI(3,4)P2, preventing the recruitment of the kinase *blk* which phosphorylates phospholipase C-gamma (PLC gamma). The production of inositol (1,4,5)-tri-phosphate (IP3) is then blocked. It leads to the inhibition of the influx of extracellular calcium, an early event triggered by the recruitment of activating receptors. It also inhibits late events such as cytokine production or cell proliferation (reviewed in O. Malbec et al., in "Immunoreceptor Tyrosine-based Inhibition Motifs", M. Daëron & E. Vivier eds, 1999, Springer-Verlag, Berlin, pp.13-27).

The biological significance of FcgammaRII-dependent negative regulation has been examined by a number of authors. It has been shown that negative regulation of antibody responses can be achieved through the recruitment of FcgammaRIIB, although it is likely that other mechanisms concur in the negative regulation of antibody responses. Thus, one can suggest that antibody-based auto-immune diseases or unwanted allo-immunization leading to the appearance of pathogenic antibodies could be circumvented through an optimized recruitment of FcgammaRIIB by antibodies or other molecules capable of binding to FcgammaRIIB.

In addition, the recruitment of FcgammaRIIB present on mast cells and basophils by allergen-complexed IgG and the subsequent co-aggregation of FcgammaRIIB with FcepsilonRI, once IgE are complexed to the same allergen, is responsible for the blockade of the anaphylactic response by these cells. Thus, IgG antibodies with an optimized ability to engage FcgammaRIIB and, hence, to trigger FcgammaRIIB inhibitory functions could represent efficient therapeutic tools in the treatment of allergy.

Finally, the inhibitory functions triggered by the recruitment of FcgammaRIIB could be exploited in cancer patients. On the one hand, it has been shown that FcgammaRIIB K.O. mice exhibit better anti-tumor responses in models where immuno-deficient FcgammaRIIB^{-/-} mice were tumor-engrafted and treated with recombinant monoclonal antibodies such as anti-CD20 or anti-HER2/Neu chimaeric or humanized antibodies, respectively. Thus, monoclonal IgG antibodies unable to trigger strong FcgammaRII inhibitory functions might

represent optimized tools in tumor treatments. On the other hand, FcγRIIb negatively regulate hematopoietic cell proliferation dependent on Receptor Tyrosine Kinases (RTKs). Bifunctional molecules such as anti-RTK IgG antibodies with optimized Fc region capable of efficiently co-aggregating RTKs with FcγRIIb could therefore block the proliferation of FcγRIIb⁺ tumor cells.

The impact of glycosylation on FcγRII/human IgG interaction has also been documented using soluble FcγRI ectodomains. Both for mouse FcγRI and human FcγRIIb, a 1:1 stoichiometry has been deduced from sedimentation equilibrium technique with ultracentrifugation and isothermal titration calorimetry (ITC) respectively (Mimura et al., J. Biol. Chem., 276, 45539-45547, 2001; Kato et al., J. Mol. Biol., 295, 213-224, 2001). Interactions between Asn²⁹⁷ and the primary GlcNAc residue were shown to be important for recognition of human chimeric IgG3 by human FcγRII (Lund et al., FASEB J., 116, 115-119, 1995). Replacement of contact residue for galactose on the alpha(1-6) mannose arm does not affect FcγRII recognition while replacement of Asp²⁶⁵, a contact for a core GlcNAc residue, results in a loss of FcγRII recognition (Jefferis et al., Immunol. Letters, 44, 111-117, 1995). In addition, another study indicated that GlcNAc residues contribute only slightly to receptor binding of human IgG1, whereas removal of alpha(1-3)- and alpha(1-6)-arm mannose residues results in a significantly decreased affinity (Mimura et al., J. Biol. Chem., 276, 45539-45547, 2001). The same study concluded that the truncation of oligosaccharides causes a close disposition of the two CH2 domains and is accompanied by a decreased binding to FcγRIIb. By contrast to FcγRIII, antibodies produced in CHO-Lec13 cells, thus devoided of fucose, showed only a slight improvement in binding to the soluble immobilized Arg¹³¹-FcγRIIA polymorphic form and to the soluble FcγRII form, and none to the soluble His¹³¹-FcγRIIA polymorphic form (both of them corresponding to the extracellular and transmembrane domains). Since the former receptors have arginine at position 131, it was postulated that the fucose may interact directly with the FcγRII residue at this position or alter the IgG1 conformation so that a slight negative effect on FcγRII binding is induced by its presence (Shields et al., J. Biol. Chem., 277,

26733-26740, 2002). The same study suggested that the galactose content does not affect binding to FcgammaRII.

5 Here, we propose to introduce changes in the glycosylation or to purify adequate glycoforms of human IgG produced for therapeutic use so as to prepare antibodies capable of exerting a fine tuning of FcgammaRIIB immuno-regulatory negative functions for example. Such method allows for the first time to identify suitable cell lines for producing antibodies with enhanced efficacy and specificity. Purification of different antibody glycoforms relies on the use of immobilized soluble forms of FcgammaR that makes it possible to enrich for
10 antibodies with an optimized or a decreased ability to bind and recruit FcgammaR.

DISCLOSURE OF THE INVENTION

15 The present invention provides a method for producing and selecting human recombinant antibodies or chimaeric antibodies or humanized antibodies or molecules that comprise Fc region of human IgG with defined glycosylation patterns by binding assays - immunofluorescence and ELISA - that use different FcR, in particular FcgammaRIIB as well as FcgammaRIII. For example, this method can be applied for selecting human recombinant antibodies or antibody-derived molecules with defined glycosylation patterns,
20 capable of specifically triggering inhibitory functions through the human type II receptors for the Fc region of IgG (FcgammaRII/CD32) by functional assays such as calcium mobilization assay and cytokine secretion assay.

25 The invention is based on the production of recombinant human, humanized or chimaeric antibodies with different glycosylation patterns by various cell lines cultured in defined media leading to specialized and fine-tuned antibodies.

In a first aspect, the invention is directed to a method for the preparation of human, humanized or chimæric antibodies or polypeptides comprising Fc region of human IgG with

enhanced therapeutic properties and decreased side effects, wherein said method comprise the step consisting of:

- a) providing candidate human, humanized or chimæric antibodies or polypeptide comprising Fc region of human IgG produced from different cell lines selected from hybridoma, heterohybridoma, animal cell lines or eukaryote microorganisms, transfected with a vector comprising the coding sequence for said antibody or polypeptide,
- b) testing the binding of said antibodies or polypeptides on Fcgamma receptors including CD16 (FcgammaRIII) and CD32 (FcgammaRIIA and B),
- c) selecting antibodies or polypeptides which :
 - i) bind specifically or preferentially to a first Fcgamma receptor,
 - ii) bind to both a first Fcgamma receptor and a second Fcgamma receptor,
 - iii) do not bind or bind only weakly to both a first Fcgamma receptor and a second Fcgamma receptor, or
 - iv) bind specifically or preferentially to said second Fcgamma receptor.

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For example, step c) may consist of selecting antibodies or polypeptides which:

- i) bind specifically or preferentially to CD16,
- ii) bind to both CD16 and CD32,
- iii) do not bind or bind only weakly to both CD16 and CD32, or
- iv) bind specifically or preferentially to CD32.

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The expression "Fc gamma receptors" refers to all Fc receptors family, including FcRγIIA, FcγRIIB, FcγRIIIA, FcgammaRIIIB and FcγRIA, FcγRIB and FcγRIC.

- 25 For example, the binding assays can be performed using FcgammaRIII and FcgammaRIIA or FcgammaRIII, FcgammaRIIA and FcgammaRIIB.

The binding assays can be performed using :

- i) indicator cells from cell lines that express different Fc receptors on their cell surface,
- ii) recombinant Fc receptors comprising FcgammaR ectodomains, Fc receptors derived-peptides or in an assay reaction mixture comprising a soluble Fc receptors or fragment thereof.

For example, one of the Fc receptors can be FcgammaRIIB and another is FcgammaRIII.

The binding assay can be followed with a functional assay. In this regard, the subset of antibodies or polypeptides selected in step c) for the feature (iv) or (ii), more particularly the subset of antibodies or polypeptides selected to bind specifically or preferentially CD32 (iv) or both CD16 and CD32 (ii), are further tested and selected for their ability to trigger or not the inhibitory functions through the human type II receptors for the Fc region of IgG (FcγRIIB/CD32).

The subset of antibodies or polypeptides selected in step c) for the feature (iv) or (ii), more particularly the subset of antibodies or polypeptides selected to bind specifically or preferentially CD32 (iv) or both CD16 and CD32 (ii) can be further tested and selected for their ability to FcγRIII/CD16 leading to improved ADCC, increased production of cytokines such as Interleukin-2 (IL-2), and/or pro-inflammatory molecules such as Tumor Necrosis Tumor alpha (TNFalpha).

Antibodies of particular interest are those who are able to induce ADCC through the FcgammaRIII and trigger the inhibitory functions through the FcgammaRIIB/CD32.

The functional assays may consist of a calcium mobilization assay and/or a cytokine secretion assay (see FR 0211416 (LFB), the method of which is incorporated herein in the description).

The functional assays may further comprise a specific FcgammaRIII ADCC assay (see WO 01/77181 (LFB), the method of which is incorporated herein in the description).

Basically, this specific FcgammaRIII ADCC assay consists of the addition of each antibody or polypeptide to a distinct reaction mixture comprising target cells, effector cells expressing FcγRIII and polyvalent IgG, and the determination of target cells lysis percentage.

Preferably, antibodies or polypeptides according to the invention are selected as displaying one of the following features:

- i) bind specifically or preferentially to CD16,
- 10 ii) bind to both CD16 and CD32,
- iii) do not bind or bind only weakly to both CD16 and CD32, or
- iv) bind specifically or preferentially to CD32.

They may be further tested in different culture conditions and different media for improving yield, antibodies or polypeptide properties as featured in i), ii), iii), or iv) and functions. No
15 binding embraces herein weak binding.

Advantageously, the invention concerns the method as depicted above, wherein antibodies or polypeptides comprising Fc region of human IgG are selected for at least one (for example two) of the following features:

20 a) giving a percentage of labeled cells (% positive cells) that represents at least a thirty-fold increase, more preferably a forty-fold increase, as compared to the percentage of cells labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD16 (FcgammaRIIIA) positive indicator cells. An example of this is the R297 antibody.
25 Another example is the 4B12 antibody.

b) giving a percentage of labeled cells (% positive cells) that represents at least a ten-fold increase, more preferably a twenty-fold increase, as compared to the percentage of cells
30 labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab)

(clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIB) positive indicator cells. An example of this is the R297 antibody. Another example is the 4B12 antibody.

- 5 c) giving a percentage of labeled cells (% positive cells) that represents at least a five-fold decrease, more preferably a ten-fold decrease, as compared to the percentage of cells labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD16 (FcγRIIA) positive indicator cells. An example of this is the AD1 antibody. Another
10 example is the B11 antibody.

- d) giving a percentage of labeled cells (% positive cells) that represents at least a three-fold decrease, more preferably a five-fold decrease, as compared to the percentage of cells labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to
15 CD32 (FcγRIIB) positive indicator cells. An example of this is the AD1 antibody. Another example is the B11 antibody.

Advantageously, the invention concerns the method as depicted above, wherein antibodies
20 or polypeptides comprising Fc region of human IgG are selected for at least one (for example two) of the following features:

- e) giving a mean fluorescence intensity (MFI) value that represents at least a 10-fold increase, more preferably a 15-fold increase, as compared to the MFI value obtained with
25 equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD16 (FcγRIIA) positive indicator cells. An example of this is the R297 antibody. Another example is the 4B12 antibody.

f) giving a mean fluorescence intensity (MFI) value that represents at least a 2-fold increase as compared to the MFI value obtained with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIb) positive indicator cells. An example of this is the R297 antibody. Another example is the 4B12 antibody.

g) a mean fluorescence intensity (MFI) value that represents at least a five-fold decrease, as compared to the MFI value obtained when equivalent doses of anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), is tested in an immunofluorescence assay that measures the binding of antibody to CD16 positive indicator cells. An example of this is the AD1 antibody. Another example is the B11 antibody.

h) a mean fluorescence intensity (MFI) value that represents at least a five-fold decrease as compared to the MFI value obtained when the same dose of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), is tested in an immunofluorescence assay that measures the binding to CD32 (FcγRIIb) positive indicator cells. An example of this is the AD1 antibody. Another example is the B11 antibody.

Advantageously, the invention concerns the method as depicted above, wherein antibodies or polypeptides comprising Fc region of human IgG are selected for one of the following features:

- an elevated mean fluorescence intensity (MFI) and a strong percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD16 (FcγRIIb) positive indicator cells. An example of this is the R297 antibody. Another example is the 4B12 antibody.

- a low mean fluorescence intensity (MFI) and a low percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD16 (FcγRIIb)

positive indicator cells. An example of this is the AD1 antibody. Another example is the B11 antibody.

5 - an elevated mean fluorescence intensity (MFI) and a strong percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD32 (FcgammaRIIB) positive indicator cells. An example of this is the R297 antibody. Another example is the 4B12 antibody.

10 - a low mean fluorescence intensity (MFI) and a low percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD32 (FcgammaRIIB) positive indicator cells. An example of this is the AD1 antibody. Another example is the B11 antibody.

15 The invention also embraces the purification of antibody glycoforms exhibiting binding properties to FcgammaRIII and FcgammaRII such as those described above by using immobilized soluble forms of FcgammaR (termed FcgammaR-immunoadsorbents) comprising the extracellular regions of the receptors. It makes it possible to enrich for antibodies with an increased or a decreased ability to bind and recruit FcgammaR. Antibodies that efficiently bind FcgammaRIII and/or FcgammaRII can be retained on such
20 FcgammaR-immunoadsorbents, subsequently eluted, allowing the obtention of preparations containing "high" FcgammaR binders antibodies. Conversely, antibodies that poorly bind FcgammaRIII and/or FcgammaRII or that do not bind FcgammaRIII and/or FcgammaRII can be obtained in the effluent fraction of FcgammaR-immunoadsorbents, allowing the obtention of preparations containing "low" FcgammaR binders antibodies.

25

As referred herein the identified antibodies or polypeptides encompasses the identified cells lines capable of producing said antibodies or polypeptides which display the above relevant features. The invention is directed to a composition comprising at least 80%, 90%, preferably at least 95%, or 99% of antibodies obtainable by the method as depicted above.

30

Therefore, in another aspect, the invention is aimed at a composition comprising at least 80%, 90%, preferably at least 95% or 99% of antibodies displaying at least one (for example 2) of the following features:

- 5 a) a MFI higher than 500, more preferably 700, and a percentage of labeled cells higher than 90 %, more preferably 95 %, in an immunofluorescence assay that measures the binding of 5, 10, 25 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD16 (FcgammaRIII) positive indicator cells (characterized by a number of FcgammaRIII receptors lower than 10^5 /cell). An example of this is the R297 antibody.
10 Another example is the 4B12 antibody.
- b) a MFI lower than 80, more preferably 20, and a percentage of labeled cells lower than 5 % , more preferably 2 %, in an immunofluorescence assay that measures the binding of 5, 10, 25 micrograms/ml antibody (containing less than 2% aggregates as determined by
15 HPLC) to 5×10^5 CD16 (FcgammaRIII) positive indicator cells (characterized by a number of FcgammaRIII receptors lower than 10^5 /cell). An example of this is the AD1 antibody.
Another example is the B11 antibody.
- c) MFI higher than 50, more preferably 100, and a percentage of labeled cells higher than 80
20 %, more preferably 90 % in an immunofluorescence assay that measures the binding of 10, 25, 50 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD32 (FcgammaRIIB) positive indicator cells (characterized by a number of FcgammaRIIB receptors lower than 5×10^4 /cell). An example of this is the R297 antibody. Another example is the 4B12 antibody.
- 25 d) a MFI lower than 50, more preferably 20, and a percentage of labeled cells lower than 5 %, more preferably 2 %, in an immunofluorescence assay that measures the binding of 10, 25, 50 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD32 (FcgammaRIIB) positive indicator cells (characterized by a number

of FcγRIIB receptors lower than $5 \times 10^4/\text{cell}$). An example of this is the AD1 antibody. Another example is the B11 antibody.

5 The invention is directed to a composition comprising at least 80%, 90 %, preferably at least 95% or 99% of antibodies or peptides displaying the following feature(s) as defined above:

- c);
- d);
- a) and c);
- b) and c);
- 10 - b) and d); or
- a) and d).

The invention also embraces the use of a cell line identified in connection with the method as defined above for producing these compositions.

15

In a still another aspect, the invention relates to the use of a composition as described in c), or b) and c), or a) and c) above to manufacture a medicament for treating auto-immune diseases or other antibody-related pathologies such as allo-immunization.

20 The invention also relates to the use of a composition as described in c), or b) and c), or a) and c) above to manufacture a medicament for treating allergies and IgE-dependent Type I hypersensitivity.

25 The invention is also directed to the use of a composition as described in c), or b) and c), or a) and c) above to manufacture a medicament for treating allergies.

Allergies include but are not limited to asthma, allergic rhinitis, allergic sinusitis, anaphylactic syndrome, urticaria, angioedema, atopic dermatitis, allergic contact dermatitis and erythema.

Alternatively, the invention encompasses the use of a composition as described in d), a) and d) or b) and d) above to manufacture a medicament for treating cancer.

5 The invention is also directed to the use of a composition as described in in c), or b) and c) or a) and c) above to manufacture a medicament for the regulation of hematopoietic cell proliferation dependent on Receptor Tyrosine Kinases (RTKs) by FcgammaRIIB. For example, said antibodies are bifunctional molecules such as anti-RTK IgG antibodies with optimized Fc region capable of efficiently co-aggregating RTKs with FcgammaRIIB, and are used to manufacture a medicament for blocking the proliferation of FcgammaRIIB tumor cells.

10 The invention further relates to the use of a composition as featured above to manufacture a medicament, wherein the antibody recognizes antigens expressed on or bound to the cell surface of target cells. Said antigens include but are not limited to allo-antigens, transplantation antigens, self-antigens such as class I and class II Major Histocompatibility Antigen (HLA), FcgammaRI, FcgammaRIIA, FcgammaRIII molecules, FcalphaRI, FcepsilonRI, B cell receptor, T cell receptor, tumor antigens such as CD20, Her2/NEU, CEA, GD2, allergen such as phospholipase A2, and IgE.

20 More particularly, the antibodies of the invention recognize Rhesus D antigen to manufacture a medicament for treating or preventing Rhesus alloimmunization of R negative patients, leading to the Hemolytic Disease of the New Born (HDNB).

25 More particularly, the antibodies of the invention recognize Rhesus D antigen to manufacture a medicament for treating or preventing Idiopathic Thrombocytopenic Purpura (ITP).

30 More particularly, the antibodies of the invention recognize HLA Class II molecules to manufacture a medicament for treating or preventing cancers, auto-immune diseases, or graft rejections.

Particular embodiments:

5 The present invention is to produce and select antibodies with increased or decreased capacity to exert immunomodulatory inhibitory functions through FcgammaRIIB. It relates to the capacity of such antibodies to negatively immunoregulate cells from the immune system or tumor cells, or not. Such a capacity is of major importance in a number of diseases and in various clinical situations such as transplantation, allo-immunization or antibody-based treatments.

10 More specifically, the present invention relates to the production and selection of antibodies with specific structural characteristics determined by different glycosylation patterns allowing them to turn on or to turn off the negative regulation exerted by FcgammaRIIB on FcgammaRIIB⁺ cells, such as B cells, monocytes or other antigen-presenting cells (APC),
15 mast cells, basophils, or any other FcgammaRIIB⁺ cells such as transformed and tumor cells.

The present invention relates to optimized antibodies that will be useful therapeutic tools for controlling antibody responses. It allows to produce and to select antibodies or other molecules optimized for binding to FcgammaRIIB, capable of triggering
20 immunomodulatory inhibitory functions through the recruitment of FcgammaRIIB in antibody-based auto-immune diseases or in unwanted allo-immunization leading to the appearance of pathogenic antibodies, notably against infused recombinant antibodies.

25 The present invention relates to optimized antibodies that will be useful therapeutic tools for controlling antibody efficacy in antibody-based treatments. It allows to produce and select antibodies with weak or no FcgammaRIIB binding, thus preventing the triggering of immunomodulatory inhibitory functions through FcgammaRIIB during antibody-based therapeutic treatments such as cancer treatments with therapeutic anti-tumor antibodies.

The present invention relates to optimized antibodies that will be useful therapeutic tools for controlling the recruitment of FcγRIIB present on mast cells and basophils by allergen-complexed IgG. As an example, the present invention allows to produce and select anti-allergen IgG antibodies with an optimized ability to engage FcγRIIB and to trigger FcγRIIB inhibitory functions through the subsequent co-aggregation of FcγRIIB with FcεRI, once IgE are complexed to the same allergen. Anti-allergen IgG antibodies with an optimized ability to engage FcγRIIB and, hence, to trigger FcγRIIB inhibitory functions could represent efficient therapeutic tools in the treatment of allergy. As an example, the present invention allows to produce and select anti-FcεRI IgG antibodies with an optimized ability to engage FcγRIIB and to trigger FcγRIIB inhibitory functions through the subsequent co-aggregation of FcγRIIB with FcεRI. Anti-FcεRI IgG antibodies with an optimized ability to engage FcγRIIB and, hence, to trigger FcγRIIB inhibitory functions could represent efficient therapeutic tools in the treatment of allergy.

The present invention relates to optimized antibodies that will be useful therapeutic tools for triggering the regulation of hematopoietic cell proliferation dependent on Receptor Tyrosine Kinases (RTKs) by FcγRIIB. Bifunctional molecules such as anti-RTK IgG antibodies with optimized Fc region capable of efficiently co-aggregating RTKs with FcγRIIB could block the proliferation of FcγRIIB⁺ tumor cells.

The present invention relates to optimized antibodies that will be useful therapeutic tools for controlling the effect of FcγRIIB expressed by APC. Examples of such effects include regulation of antigen presentation and T cell recruitment and regulation of acquired immunity.

Binding to FcγRIII as used herein refers to a binding of antibodies where only a low percentage of aggregates is present in antibody preparation (<2%), and where antibodies are not complexed on purpose by any mean such as heat or chemical aggregation or by formation of immune-complexes (IC) with the relevant antigen.

Binding to FcgammaRIII as used herein refers to a binding of antibodies complexed with F(ab)₂ anti-human Ig or under immune-complexes (IC) forms with the relevant antigen.

5 Binding to FcgammaRIIB as used herein refers to a binding of antibodies where only a low percentage of aggregates is present in antibody preparation (<2%), and where antibodies are not complexed on purpose by any mean such as heat or chemical aggregation or by formation of immune-complexes (IC) with the relevant antigen.

10 Binding to FcgammaRIIB as used herein refers to a binding of antibodies complexed with F(ab)₂ anti-human Ig or under immune-complexes (IC) forms with the relevant antigen.

15 An activity of antibodies of the present invention includes the inhibition of their binding to FcgammaRIII expressed on indicator cells by antibodies directed to FcgammaRIII.

An activity of antibodies of the present invention includes the inhibition of their binding to FcgammaRIIB expressed on indicator cells by antibodies directed to FcgammaRII.

20 An activity of antibodies of the present invention includes the inhibition of calcium mobilization, by co-aggregation of such antibodies bound to surface FcgammaRIIB with activating surface receptors such as BCR, TCR, FcepsilonRI, or not. Calcium mobilization as used herein refers to the influx of extracellular calcium into a cell following the recruitment of activating receptors such as BCR.

25 An activity of antibodies of the present invention includes the inhibition of cytokine production by co-aggregation of such antibodies bound to surface FcgammaRIIB with activating surface molecules such as BCR, or not.

Cytokine production as used herein refers to the release of cytokine by a cell following the recruitment of activating receptors such as BCR.

5 The inhibition of cytokine production by molecules of the present invention include inhibition of Interleukin-2 (IL-2) production.

The method of the present invention for selecting an immunologically active molecule described below using human IgG antibodies directed against RhD antigen and chimeric mouse/human IgG antibodies directed against HLA-DR class II molecules. As example
10 R297 and AD1 human monoclonal antibodies directed against Rhesus D and 4B12 and B1 chimeric mouse/human antibodies directed against HLA-DR are being used. R297 and 4B12 antibodies are produced in YB2/0 cells (ATCC number CRL-1662). B11 antibody produced by wild-type CHO-DG44 cells. AD1 antibody is produced by cells derived from human B lymphocyte from a donor with anti-RhD antibodies fused with the heteromyeloma
15 cell line K6H6/B5 (ATCC number CRL-1823).

The capacity of the purified human or chimeric antibody, either complexed with F(ab)₂ anti-human Ig or not, to bind FcγRIIB1 expressed on indicator cells (mouse IIA1.6 lymphoma B cells expressing a functional mouse BCR and a human recombinant FcγRIIB1, termed IIA1.6-huFcγRIIB1) can be measured by a fluorescence
20 antibody assay.

The capacity of the purified human or chimeric antibody to induce the inhibition of calcium mobilization can be assessed by a calcium measurement assay. Inhibition of calcium
25 mobilization is evaluated by co-aggregating the purified human or chimeric antibody bound to FcγRIIB1 with BCR expressed on indicator cells (mouse IIA1.6 lymphoma B cells expressing a functional mouse BCR and a human recombinant FcγRIIB1). It is compared to BCR-mediated calcium mobilization induced with F(ab)₂ rabbit anti-goat IgG (RAG) that cross react with mouse IgG.

The capacity of the purified human or chimeric antibody to induce the inhibition of interleukin-2 (IL-2) production can be measured by an enzyme-linked immunosorbent assay (ELISA) that detects and quantifies IL-2. Inhibition of IL-2 production is evaluated by co-aggregating the purified human or chimeric antibody bound to FcgammaRIIB with BCR
 5 expressed on indicator cells (mouse IIA1.6 lymphoma B cells expressing a functional mouse BCR and a human recombinant FcgammaRIIB1). It is compared to BCR-mediated IL-2 production induced with F(ab)₂ goat anti-mouse IgG (H+L).

Further details are given below in example 1. Results of these binding and functional tests
 10 are presented in the figures.

EXAMPLE 1: Selection of anti-RhD antibodies with improved properties to trigger FcgammaRIIB mediated inhibition.

15 1. FIGURE LEGENDS, RESULTS AND COMMENTS

Figure 1 represents HPLC profiles of different monoclonal antibodies in order to control the purity and the integrity of the molecules and the absence of polymerized forms : R297 and AD1 anti-RhD antibodies, 4B12 and B11 anti-HLA-DR antibodies.

20

Name	Retention time (min)	Area	% Area
<u>Peak 1</u>	28.300	72387414	99.04
<i>Peak 2</i>	45.267	700356	0.96

R-297 monoclonal antibody is composed of 99% of monomers.

25

Figure 2 represents the glycosylation patterns of different antibodies, as determined by capillary electrophoresis. The results are expressed as percentage of the different structures identified. The antibodies were expressed in different cell lines : R297 anti-RhD in YB2 cell line, AD1 anti-RhD in an heterohybridoma cell line, 4B12 anti-HLA-DR in YB2/0 cell line and B11 anti-HLA-DR in CHO-DG44 cell line.

Figure 3 represents the binding of F(ab)₂ goat anti-human IgG (H+L)-complexed R297 and AD1 antibodies to human FcγRIIB1 expressed on IIA1.6 cells *as described in [32]*. The binding of R297 and AD1 human anti-RhD antibodies to human FcγRIIB1 is assessed by indirect immunofluorescence. Human antibodies (1μg/ml, 5μg/ml, 10μg/ml, 25μg/ml ; 50μg/ml) are first complexed with F(ab)₂ goat anti-human IgG (H+L) (at doses of 1.5μg/ml, 7.5μg/ml, 15μg/ml, 40μg/ml, and 80μg/ml, respectively) and then incubated with IIA1.6-huFcγRIIB1 cells. Binding of R297 and AD1 antibodies is then revealed with FITC labelled-F(ab)₂ mouse anti-human IgG (H+L) (black curves). The dotted curves show background fluorescence of cells incubated with FITC-F(ab)₂ mouse anti-human IgG (H+L) only.

Results and comments : complexed R297 antibody significantly binds to IIA1.6 huFcγRIIB1 cells even at doses as low as 5μg/ml and 10μg/ml. At these doses, MFI are 111 and 117, respectively, with more than 70% labelled cells. At higher doses, up to 98% cells are stained, with MFI reaching 327 at 50μg/ml. By contrast, no binding of AD1 antibody to human FcγRIIB1 expressed by IIA1.6 cells is observed at a dose of 10μg/ml. Only high doses of complexed AD1 antibody bind very weakly to IIA1.6 huFcγRIIB1 cells. Only 9% of FcγRIIB1⁺ IIA1.6 cells are weakly labeled (MFI = 46) at 50μg/ml. Last, no binding on FcγRIIB negative parental IIA1.6 cells is observed with any of the antibodies (not shown). Thus, R297, produced by YB2/0 cells, with a well defined glycosylation pattern, is an example of a FcγRIIB “high binder” monoclonal antibody, whereas AD1, produced by a cell line obtained by fusing human B cells with heteromyeloma cells (K6H6 x B5), also with a well-defined glycosylation pattern, is an example of a FcγRIIB “low binder” monoclonal antibody.

Figure 4 represents the binding of F(ab)₂ goat anti-human IgG (H+L)-complexed 4B12 and B11 anti-HLA-DR chimeric mouse/human antibodies to human FcγRIIB1 expressed on IIA1.6 cells as described in [32]. The binding of 4B12 and B11 chimeric mouse/human anti-HLA-DR antibodies to human FcγRIIB1 is assessed by indirect immunofluorescence.

4B12 and B11 antibodies (1μg/ml, 5 μg/ml, 10μg/ml, 25μg/ml, 50μg/ml) are first complexed with F(ab)₂ goat anti-human IgG (H+L) (at doses of 1.5 μg/ml, 7.5 μg/ml, 15μg/ml, 40μg/ml, and 80μg/ml, respectively) and then incubated with IIA1.6-huFcγRIIB1 cells. 4B12 and B11 antibody binding is revealed with FITC labelled-F(ab)₂ mouse anti-human IgG (H+L) (black curves). The dotted curves show background fluorescence of cells incubated with FITC-F(ab)₂ mouse anti-human IgG (H+L) only.

Results and comments : complexed 4B12 antibody significantly binds to IIA1.6-huFcγRIIB1 cells at all the doses tested. The percentage of labelled cells is 13 % at a dose of 1 μg/ml and reaches more than 60% at 10 μg/ml. At this dose, a MFI of 103 is observed. No significant binding of B11 antibody to human FcγRIIB1 expressed by IIA1.6 cells is observed up to 25 μg/ml. Even at a dose of 50 μg/ml, only 5 % cells are labelled with a MFI of 92. Last, no binding on FcγR negative parental IIA1.6 cells is observed with any of the antibodies (not shown). Thus, 4B12, produced by YB2/0 cells, with a well-defined glycosylation pattern, is a second example of a FcγRIIB "high binder" monoclonal antibody, whereas B11, produced by CHO-Lec1 cells, also with a well-defined glycosylation pattern, is a second example of a FcγRIIB "low binder" monoclonal antibody.

Figure 5 represents the binding of non-complexed R297 and AD1 human anti-RhD antibodies to human FcγRIIB1 expressed on IIA1.6 cells as described in [32]. Less than 2% aggregates are detected in the antibody preparations. R297 and AD1 antibodies (1μg/ml, 5μg/ml, 10μg/ml, 25μg/ml, 50μg/ml) are incubated with IIA1.6-huFcγRIIB1 cells. The binding of R297 and AD1 to human FcγRIIB1 is then assessed by indirect immunofluorescence. It is revealed with FITC labelled-F(ab)₂ mouse anti-human IgG (H+L)

(black curves). The dotted curves show background fluorescence of cells incubated with FITC-F(ab)₂ mouse anti-human IgG (H+L) only.

Results and comments : non-complexed R297 antibody significantly binds to IIA1.6 huFcγRIIB1 cells even at a low dose such as 5 μg/ml (MFI : 28). Up to 86% cells are labelled at a dose of 50 μg/ml, with a MFI reaching 60. No binding of AD1 antibody to human FcγRIIB1 expressed by IIA1.6 cells is observed at any dose ranging from 1 μg/ml up to 50 μg/ml (percentage of labelled cells <2%, with a MFI of 12 that corresponds to the background fluorescence obtained with the secondary revealing FITC labelled-F(ab)₂ mouse anti-human IgG (H+L). Thus, R297 is an example of recombinant antibody produced by YB2/0 cells, with a well-characterized glycosylation pattern, that strongly binds FcγRIIB present on cell surface, even under monomeric forms. By contrast, AD1 antibody is an example of recombinant antibody, produced by a cell line obtained by fusing human B cells with heteromyeloma cells (K6H6 x B5), that does not bind significantly to membrane FcγRIIB, even when used at doses 10 fold-higher than R297 dose (example: 50 μg/ml versus 5 μg/ml in the Figure 4)

Figure 6 represents the inhibition of the human anti-RhD antibody R297 binding to human FcγRIIB1 by the mouse AT10 monoclonal antibody directed against human FcγRII. The binding of R297 to human FcγRIIB1 is assessed by indirect immunofluorescence. IIA1.6-huFcγRIIB1 cells are first incubated with the mouse mAb AT10 directed against the binding site of human FcγRIIA and FcγRIIB at doses of 0.1 μg/ml, 0.5 μg/ml, 1 μg/ml, or 5 μg/ml, and then with 25 μg/ml of R297 human anti-RhD antibody complexed with 40 μg/ml F(ab)₂ goat anti-human IgG (H+L). The binding of R297 mAb to human FcγRIIB1 is then assessed by indirect immunofluorescence using FITC-F(ab)₂ mouse anti-human IgG (H+L) as revealing antibodies (black curves). The dotted curves show background fluorescence of cells incubated with FITC-F(ab)₂ mouse anti-human IgG (H+L) only.

Results and comments : a dose-dependent inhibition of the binding of R297 mAb to surface FcγRIIB is achieved when the mAb AT10 directed against the binding site of FcγRIIB is used as competitor. As little as 0.1 μg/ml AT10 allows to decrease the MFI

down from 420 to 232. When 5 $\mu\text{g/ml}$ AT10 are used, only a marginal binding of R297 to FcgammaRIIB IIA1.6 cells is observed (% labeled cells: 11%, MFI: 59). It indicates that the binding of R297 antibody to FcgammaRIIB⁺ IIA1.6 cells is mediated through the FcgammaRIIB expressed on these cells.

5

Figure 7 represents the inhibition of the human anti-RhD antibody R297 binding to FcgammaRIIB1⁺ IIA1.6 cells by F(ab)₂ mouse anti-human FcgammaRII mAb, 6C4.

IIA1.6-huFcgammaRIIB1 cells are first incubated with F(ab)₂ 6C4 mAb directed against the binding site of human FcgammaRII at doses of 1 $\mu\text{g/ml}$, 10 $\mu\text{g/ml}$, 25 $\mu\text{g/ml}$, 50 $\mu\text{g/ml}$ or 100 $\mu\text{g/ml}$, and then with 25 $\mu\text{g/ml}$ of R297 complexed with 40 $\mu\text{g/ml}$ F(ab)₂ goat anti-human IgG (H+L). The binding of R297 mAb to human FcgammaRIIB1 is then assessed by indirect immunofluorescence using FITC-F(ab)₂ mouse anti-human IgG(H+L) as revealing antibodies (black curves). The dotted curves show background fluorescence of cells incubated with FITC-F(ab)₂ mouse anti-human IgG (H+L) only.

Results and comments : a dose-dependent inhibition of the binding of R297 mAb to surface FcgammaRIIB is achieved when the F(ab)₂ 6C4 mAb directed against the binding site of FcgammaRIIB is used as competitor. 10 $\mu\text{g/ml}$ F(ab)₂ 6C4 allow to decrease the MFI down from 420 to 131. When 100 $\mu\text{g/ml}$ F(ab)₂ 6C4 are used, only a marginal binding of R297 to FcgammaRIIB IIA1.6 cells is observed (% labeled cells: 7%, MFI: 68). This dose-dependent inhibition obtained with a F(ab)₂ fragment of a monoclonal antibody directed against the FcgammaRII binding site indicates that the binding of R297 antibody to FcgammaRIIB⁺ IIA1.6 cells is mediated through the FcgammaRIIB expressed on these cells.

Figure 8 represents the inhibition of the binding of the chimeric mouse/human anti-HLA-DR antibody 4B12 to FcgammaRIIB1⁺ IIA1.6 cells by mouse AT10 monoclonal antibody and by mouse F(ab)₂ 6C4 monoclonal antibody, both directed against the binding site of human FcgammaRII. IIA1.6-huFcgammaRIIB1 cells are first incubated with AT10 (A) or F(ab)₂ 6C4 (B) mAbs at doses of 0.1 $\mu\text{g/ml}$, 0.5 $\mu\text{g/ml}$, 1 $\mu\text{g/ml}$ for AT10, or 1 $\mu\text{g/ml}$, 10 $\mu\text{g/ml}$, 25 $\mu\text{g/ml}$ for F(ab)₂ 6C4. IIA1.6-huFcgammaRIIB1 cells are then incubated with 25 $\mu\text{g/ml}$ of 4B12 complexed with 40 $\mu\text{g/ml}$ F(ab)₂ goat anti-human IgG (H+L). The binding

of 4B12 mAb to human FcγRIIB1 is then assessed by indirect immunofluorescence using FITC-F(ab)₂ mouse anti-human IgG(H+L) as revealing antibodies (black curves). The dotted curves show background fluorescence of cells incubated with FITC-F(ab)₂ mouse anti-human IgG (H+L) only.

- 5 Results and comments : a strong inhibition of the binding of 4B12 mAb to surface FcγRIIB is achieved when the AT10 and the F(ab)₂ 6C4 mAbs directed against the binding site of FcγRIIB are used as competitors. One μg/ml F(ab)₂ 6C4 leads to 50% inhibition of 4B12 binding (% labelled cells : from 43% to 22% ; MFI: from 113 to 76). 10μg/ml F(ab)₂ 6C4 totally inhibits the binding of 4B12 (percentage of labelled cells 2% down from 43% ; MFI down from 113 to 36). Similarly, a strong inhibition of the binding of 4B12 mAb to surface FcγRIIB is achieved even at low doses of AT10 mAb. As little as 0.1μg/ml AT10 allows to decrease the MFI down from 113 to 76 and the percentage of cells down from 43% to 22%. When 0.5μg/ml AT10 is used, only a marginal binding of 4B12 to FcγRIIB IIA1.6 cells is observed (% labeled cells: 3%, MFI: 109). 1μg/ml AT10 completely blocks the binding of 4B12 to FcγRIIB IIA1.6 cells (MFI: 12 ; % labelled cells: <2%). Thus, as observed with the anti-RhD 297 antibody, the binding of the anti-HLA-DR 4B12 antibody, produced by YB2/0 cells, to FcγRIIB IIA1.6 cells is mediated through the FcγRIIB expressed on these cells. It indicates that recombinant antibodies produced by YB2/0 cells exhibit a strong binding to FcγRIIB expressed on surface cells, whatever their specificities.

Figure 9 represents a comparison of the binding of human R297 and AD1 anti-RhD monoclonal antibodies to human FcγRIIIA expressed by stably transfected Jurkat cells (Jurkat-huFcγRIIIA/γ) with the binding of the chimeric mouse/human anti-CD20 Rituxan (rituximab) monoclonal antibody. In these experiments, antibodies are not aggregated and are not bound to their specific antigen. 5 x 10⁵ Jurkat-huFcγRIIIA/γ cells are first incubated with the various mAbs used at doses of 5μg/ml, 10μg/ml or 25μg/ml. The binding of monoclonal antibodies to human FcγRIIIA is then assessed by indirect immunofluorescence using FITC-F(ab)₂ mouse anti-human IgG(H+L) as revealing antibodies. Flow cytometry acquisition is performed with a FACScan Calibur.

(Becton Dickinson, Mountain View, California, USA), and analysis is performed with the Cell Quest Pro software. Percentage of positive cells (A) and Mean Fluorescence Intensity (MFI) (B) are calculated after acquisition and analysis of ten thousand events for each condition. The value of background MFI obtained with the revealing FITC-F(ab)₂ mouse anti-human IgG(H+L) antibodies is 12.

Results and comments : Panel A and panel B show that three patterns of binding are observed depending on the monoclonal antibody tested. The anti-RhD R297 monoclonal antibody binds more than 90% of FcγRIIIA positive Jurkat cells whatever the doses used (5μg/ml, 10μg/ml and 25μg/ml). It is considered therefore as a high binder, with a MFI ranging from around 500 up to 700 (panel B). By contrast, the anti-CD20 monoclonal antibody Rituxan binds only weakly to FcγRIIIA positive Jurkat cells ; around 30% cells are labeled when the Rituxan antibody is used at the higher dose (25μg/ml) ; only 10% cells are labeled with 10μg/ml of Rituxan, while there is no significant labeling when the antibody is tested at 5μg/ml (panel A). In addition, a low MFI is observed whatever the dose of Rituxan tested (i.e., <100) (panel B). Moreover, the monoclonal antibody AD1, directed against RhD, shows a third pattern of binding to FcγRIIIA Jurkat cells. This antibody binds a very low percentage of cells at any dose tested (panel A) (<5%) with a low MFI (<70) at each dose. Thus, three groups of monoclonal antibodies with Fc region from the human IgG1 subclass can be defined using this assay that uses antibody preparations with no or very few aggregates (less than 2% as determined by HPLC) : firstly, "high" mAb binders that bind a high percentage of FcγRIIIA positive cells (>90%) with an elevated MFI (>500), even at a low dose (5μg/ml) ; secondly, "intermediate" mAb binders that bind significantly FcγRIIIA positive cells only at the higher dose of antibody tested (about 30% labelled cells at 25μg/ml), whereas lower doses show only a marginal binding. These "intermediate" mAb binders label cells with a low MFI (<100) whatever the dose tested ; thirdly, "low" mAb binders that bind less than 5% cells, with a low MFI (<70), whatever the dose tested.

Figure 10 represents a comparison of the binding of human R297 and AD1 anti-RhD monoclonal antibodies to human FcγRIIB1 expressed by IIA.1.6 cells (IIA.1.6-

huFcgammaRIIB1 cells) with the binding of the chimeric mouse/human anti-CD20 Rituxa (rituximab) monoclonal antibody. In these experiments, antibodies are not aggregated and are not bound to their specific antigen. 5×10^5 IIA1.6-huFcgammaRIIB1 cells are first incubated with the various mAbs used at doses of 10µg/ml, 25µg/ml or 50µg/ml. The binding of monoclonal antibodies to human FcgammaRIIB1 is then assessed by indirect immunofluorescence using FITC-F(ab)₂ mouse anti-human IgG(H+L) as revealing antibodies. Flow cytometry acquisition is performed with a FACScan Calibur (Becton Dickinson, Mountain View, California, USA), and analysis is performed with the Cell Quest Pro software. Percentage of positive cells (A) and Mean Fluorescence Intensity (MFI) (B) are calculated after acquisition and analysis of ten thousand events for each condition. The value of background MFI obtained with the revealing FITC-F(ab)₂ mouse anti-human IgG(H+L) antibodies is 12.

Results and comments : Panel A and panel B show that three patterns of binding are observed depending on the monoclonal antibody tested. The anti-RhD R297 monoclonal antibody binds more than 50% IIA1.6-huFcgammaRIIB1 cells at a dose of 10µg/ml and more than 80% at doses of 25µg/ml and 50µg/ml. It is considered therefore as a high FcgammaRIIB binder, with a MFI reaching more than 120 at a dose of 50µg/ml (panel B). By contrast, the anti-CD20 monoclonal antibody Rituxan binds only weakly to IIA1.6 huFcgammaRIIB1 cells ; around 25% cells are labeled when the Rituxan antibody is used at the higher dose (50µg/ml) ; less than 5% cells are labeled with 25µg/ml of Rituxan, while there is no significant labeling when the antibody is tested at 10µg/ml (panel A). In addition, a low MFI is observed whatever the dose of Rituxan tested (<60) (panel B). Moreover, the monoclonal antibody AD1, directed against RhD, shows a third pattern of binding to IIA1.6 huFcgammaRIIB1 cells. This antibody does not bind the cells when tested at 10µg/ml and 25µg/ml and binds a very low percentage of cells at 50µg/ml (panel A) (<5%) with a low MFI at this dose (<65). Thus, three groups of monoclonal antibodies with Fc region from the human IgG1 subclass can be defined using this assay that uses antibody preparation with no or very few aggregates (less than 2% as determined by HPLC) : firstly, "high" mAb binders that bind a high percentage of huFcgammaRIIB1 positive cells (>80%) with an elevated MFI (>70), at doses of 25µg/ml and 50µg/ml ; secondly, "intermediate" mAb

binders that bind significantly huFcgammaRIIB1 positive cells only at the higher dose of antibody tested (about 25% labeled cells at 50µg/ml), whereas lower doses show only a marginal binding, if any. These "intermediate" mAb binders label huFcgammaRIIB1 positive cells with a low MFI (<60) whatever the dose tested ; thirdly, "low" mAb binders that do not bind huFcgammaRIIB1 positive cells at doses of 10µg/ml and 25µg/ml, and that bind less than 5% cells, with a low MFI (<70), at a dose of 50µg/ml.

Figure 11 summarizes the binding data (% labeled cells and Mean Fluorescence Intensity, MFI) obtained with the human R297 and AD1 anti-RhD monoclonal antibodies and with the chimeric mouse/human anti-CD20 Rituxan (rituximab) monoclonal antibody when FcgammaRIIIA positive Jurkat cells and IIA1.6-huFcgammaRIIB1 positive cells are tested in indirect immunofluorescence assays (see legends to Figures 9 and 10). Percentage of positive cells are represented in ordinate and MFI are represented in abscissa. Upper and lower left panels indicate % and MFI values obtained with R297, AD1 and Rituxan (rituximab) monoclonal antibodies. Upper and lower right panels indicate % and MFI values obtained with AD1 and Rituxan (rituximab) monoclonal antibodies using a different scale in ordinate and abscissa.

Results and comments: three groups of monoclonal antibodies with Fc region from the human IgG1 subclass can be defined using this indirect immunofluorescence assay with antibody preparations that contain no or very few aggregates (less than 2% as determined by HPLC) : i) antibodies that give rise to an elevated MFI and to a high percentage of labeled cells (>80%) at various doses (FcgammaRIIIA, 5, 10, 25µg/ml ; FcgammaRIIB1, 10, 25, 50µg/ml) such as the R297 monoclonal antibody (FcgammaRIIIA: upper left panel ; FcgammaRIIB1: lower left panel) ; ii) antibodies that give rise to an intermediate MFI and to an intermediate percentage of labeled cells (20-30%) only at a high dose (FcgammaRIIIA, 25µg/ml ; FcgammaRIIB1, 50µg/ml), such as the Rituxan monoclonal antibody (FcgammaRIIIA: upper right panel ; FcgammaRIIB1: lower right panel, grey symbols) ; iii) antibodies that give rise to an intermediate MFI and to a low percentage of labeled cells (<5-10%) only at a high dose (FcgammaRIIIA, 25µg/ml ; FcgammaRIIB1, 50µg/ml), such as the

AD1 monoclonal antibody (FcgammaRIIA : upper right panel ; FcgammaRIIB1 : lower right panel, open symbols).

Figure 12 represents the inhibition of calcium mobilization by R297 human anti-Rh antibody. IIA1.6-huFcgammaRIIB1 cells are stimulated either with F(ab)₂ rabbit anti-goat IgG (RAG) (10µg/ml) that cross-react with mouse Ig or with R297 antibody (100µg/ml) in presence of F(ab)₂ goat anti-human IgG (H+L) (GAH) (10µg/ml) and F(ab)₂ rabbit anti-goat IgG (RAG) (10µg/ml). This latter assay allows to efficiently cross-link murine surface IgG with the human FcgammaRIIB1 expressed by IIA1.6-huFcgammaRIIB1 cells. The Ca²⁺ influx observed in this condition of BCR/ FcgammaRIIB1 cross-linking is compared to BCR-mediated Ca²⁺ influx induced with F(ab)₂ RAG (10µg/ml). Intracellular free Ca²⁺ levels in Fluo-3 AM loaded cells are monitored by flow cytometry. Ca²⁺ release from intracellular Ca²⁺ stores is measured in presence of 1mM EGTA (left part of the curves), and Ca²⁺ entering the cells is measured after addition of 6mM CaCl₂ (right part of the curves).

Results and comments : the addition of F(ab)₂ rabbit anti-goat IgG (RAG) antibody triggers calcium release from the intracellular stores (left peak of the curve, dotted line) as well as a calcium influx into IIA1.6-huFcgammaRIIB1 cells (right peak of the curve, dotted line). A strong inhibition of the calcium influx (right peak of the curve, black line), characteristics of the engagement of FcgammaRIIB on B cells, is obtained by incubating IIA1.6-huFcgammaRIIB1 cells with R297 antibody (100µg/ml) in presence of F(ab)₂ goat anti-human IgG (H+L) (GAH) (10µg/ml) and F(ab)₂ rabbit anti-goat IgG (RAG) (10µg/ml). As expected when the FcgammaRIIB inhibitory function is triggered, no inhibition of the calcium release from intracellular stores is observed (left peak of the curve, black line). Thus, this experiment shows that the R297 monoclonal antibody is capable of triggering inhibitory functions mediated through FcgammaRIIB expressed on lymphoma B cells, triggering which is known to lead to the blockade of B cell activation and differentiation in antibody-producing cells.

Figure 13 represents the inhibition of IL-2 production by R297 but not by AD1 human anti-RhD antibodies. 5×10^5 IIA1.6-huFcgammaRIIB1 cells are stimulated for 22h either with

50µg/ml F(ab)₂ rabbit anti-goat IgG (RAG) alone or with 50µg/ml F(ab)₂ rabbit anti-goat IgG and R297 or AD1 antibody at different concentrations as indicated in abscissa (from 1 to 100µg/ml). The RAG F(ab)₂ fragments cross-react with both murine and human IgG, allowing to cross-link the mouse surface IgG expressed by IIA1.6 huFcγRIIB1 positive cells and the human monoclonal IgG antibody bound to FcγRIIB1. The production of mouse IL-2 by IIA1.6-huFcγRIIB1 cells in cell culture supernatants is measured by an ELISA assay using 1µg/ml rat anti-mouse IL-2 as capture antibody and 1µg/ml biotinylated rat anti-mouse IL-2 as detection antibody.

Results and comments: the stimulation of IIA1.6-huFcγRIIB1 positive cells that express surface IgG by F(ab)₂ rabbit anti-goat IgG (RAG) cross-reacting with mouse IgG triggers the secretion of IL-2 (about 380 pg/ml after 22h of culture of 5 x 10⁵ IIA1.6 FcγRIIB1 cells). Strikingly, the presence of various doses of R297 antibody (from 1µg/ml to 100µg/ml) in presence of RAG F(ab)₂ fragments cross-reacting with both murine and human IgG induces a dose-dependent inhibition of IL-2 secretion. As low as 1µg/ml R297 antibody induces about a 30% inhibition of IL-2 production, while 50µg/ml induces up to 50% inhibition. By contrast, AD1 antibody does not inhibit IL-2 production by IIA1.6-huFcγRIIB1 cells cultured in presence of RAG F(ab)₂ fragments, even when used at a high dose such as 100µg/ml (<3% decrease). Thus, this experiment shows that the R297 monoclonal antibody, a high mAb binder to FcγRIIB1 expressed by IIA1.6-huFcγRIIB1 cells, is capable of triggering inhibitory functions mediated through FcγRIIB expressed on lymphoma B cells. Notably, it leads to the blockade of IL-2 production by these cells. In addition, this experiment also shows that the AD1 monoclonal antibody, a low mAb binder to FcγRIIB1 expressed by IIA1.6-huFcγRIIB1 cells, is not able to trigger inhibitory functions mediated through FcγRIIB expressed on lymphoma B cells.

EXAMPLE 1: Selection of anti-RhD antibodies with improved properties to trigger FcγRIIB mediated inhibition.

Material and methods

Cell lines and antibodies

The mouse IIA1.6 B cell lymphoma is a FcγR-defective variant of A20 B cells (Jor
5 *et al.*, 1986) that bears a deletion of the 5' end of the FcγRII gene (Lewis *et al.*, 1986;
Bonnerot *et al.*, 1991) and does not transcribe the genes encoding the α- and the γ-chains
mouse FcγRIII (Bonnerot *et al.*, 1991). IIA1.6 cells were transfected by the cDNA
encoding FcγRIIB1 obtained from Dr. M. Hogarth (Melbourne University, Parkville
Victoria, Australia) (Hogarth *et al.*, 1987). IIA1.6 and IIA1.6-huFcγRIIB1 cells were
10 cultured in Click medium [RPMI 1640 (GIBCO, Paisley, Scotland) supplemented with 10
heat-inactivated FCS (Hyclone Laboratories Inc., Logan, Utah, USA), 100U/ml penicillin,
100μg/ml streptomycin, 2mM L-glutamine, 5mM sodium pyruvate, 0.5μM 2-
mercaptoethanol.

The AD1 B cell line was obtained by fusing human B lymphocytes from a donor
15 immunized with human Rh positive red blood cells with the heteromyeloma cell line
K6H6/B5 (ATCC number CRL-1823) and subsequent cloning was performed by
limiting dilutions. AD1 cells were cultured in RPMI 1640 medium supplemented with
5% FCS.

The YB2/0 cell line (ATCC number CRL-1662) was used to produce a human
20 recombinant anti-Rh D (R297) monoclonal antibody and a chimeric anti-HLA-DR
(4B12) recombinant monoclonal antibody. YB2/0 transfected cells were cultured in a
specific EM culture medium with 5% of FCS.

CHO DG44 cells (ATCC number CRL-1735) were cultured in a specific EM culture
medium supplemented with 5% of FCS to produce the chimeric anti-HLA-DR (B11)
25 recombinant antibody.

All the producing cell lines were adapted and cultured in a protein free culture medium
and the production were performed in rolling bottles or in fermentors. After the
production, the monoclonal antibodies were purified by using protein-A affinity
chromatography. The purified antibodies were then stored frozen at -20°C.

Rituxan (Rituximab) (Genentech, South San Francisco, California, USA) is a chimeric mouse/human anti-CD20 monoclonal antibody used in the treatment of patients with relapsed or refractory low-grade or follicular, CD20⁺, B-cell non -Hodgkin's lymphoma (NHL). Commercial Rituxan vials are stored at 2°C to 8°C in a sterile preservative-free solution at a concentration of 10mg/ml.

Immunofluorescence assay

First, recombinant human antibodies were complexed under polymeric forms for 30 min with affinity-purified goat F(ab)₂ anti-human IgG (Jackson ImmunoResearch Laboratories Inc., West Grove, Pennsylvania, USA). 5x10⁵ indicator cells were then incubated for 30 min with complexed antibodies in ice-cold phosphate-buffered saline containing 0.5% bovine serum albumin (PBS-BSA). Cells were then washed with PBS-BSA, and antibody binding was detected by incubation with FITC-labelled mouse F(ab)₂ anti-human IgG (H+L) (Jackson ImmunoResearch) for 30 min on ice. After further washing, flow cytometry analysis was performed with a FACScalibur 4CA (Becton Dickinson, Mountain View, California, USA), using the Cell Quest Pro software.

Facsclibur 4CA technical specifications

A 15mW, 488nm air cooled Argon-ion laser is used for acquisition of FITC fluorochrome. Estimated detection limit is 200 FITC equivalent molecules per particle. Logarithmic amplifiers for FL1 (FITC) provide four log decade range. Sorting speed is 300cells/sec and the sorting purity is more than 95%.

Calcium mobilization assay

IIA1.6-huFcγRIIB1 cells were incubated or not with R297 or AD1 mAb for 30 min on ice. 10⁶ cells were incubated with 5mM Fluo-3 AM (Molecular Probes, Eugene, Oregon, USA) for 30 min at room temperature in RPMI 1640 containing 0.2% Pluronic F-127 (Sigma Chemicals Co., St. Louis, Missouri, USA). The loaded cells were then washed three times with RPMI 1640 and adjusted to 10⁶/ml. Cells were stimulated with rabbit F(ab')₂ anti-

goat IgG (H+L) (RAG) (Jackson ImmunoResearch) alone or rabbit F(ab)₂ anti-goat IgG (H+L) (RAG) and goat F(ab)₂ anti-human IgG (H+L) (GAH) (Jackson ImmunoResearch) in RPMI 1640 containing 1mM EGTA. After 150 sec of stimulation, 6mM CaCl₂ were added. Intracellular calcium mobilization was detected by flow cytometry, performed with FACScalibur 4CA, using the Cell Quest Pro software. Intracellular calcium concentration means were then calculated with the FCS assistant 1.2.9 beta software (Becton Dickinson).

Detection of IL-2 production

10⁵ IIA1.6-huFcγRIIB1 cells were stimulated for 18 h with F(ab)₂ rabbit anti-goat (Jackson ImmunoResearch) alone or with F(ab)₂ rabbit anti-goat and R297 or AD1. The presence of mouse IL-2 was measured by ELISA assay, using 1μg/ml rat anti-mouse IL-2 (Pharmingen) as capture antibody and 1μg/ml biotinylated rat anti-mouse IL-2 (Pharmingen) as detection antibody.

15 Binding studies on soluble Fc fragments

Antibody interaction with the Fc receptor is the primary event needed for cell regulation. However, cells or cell ghosts expressing are not so easy to use “in vitro” and require either negative control (not always available) or inhibition studies, to insure the binding specificity. To overcome these problems, recombinant soluble Fc receptor were used to evaluate the binding capacity of monoclonal or polyclonal antibodies to Fc receptor. Two types of experiments were performed :

- binding studies in the absence of antigens, in this condition, the data were assessed by flow cytometry by measuring MFI
- binding studies in the presence of antigens : the data were assessed by using immunoenzymatic tests in which antigen or antigen bearing membranes were coated onto microplates.

Following described method was used for anti-D antibodies using Rhesus positive red cells. However, it could be extended to other antigens, by using the appropriate cell carrying onto its cell membrane the specific antigen.

Binding assays with anti-RhD antibodies were performed as following :

- 1- Rhesus positive red cells were treated with papaine (0.1mg/ml) for 10 minutes at 37°C. After washing, red cells were diluted in cold CFC buffer (0,1M glucose; 0,07M NaCl; 0,02M boric acid; 0,035M tri sodium citrate; 0,002M tétra-sodium EDTA).
- 5 Papaine treated red cells (10^7 per ml) were incubated for 2 minutes in a P96 microtiter U well plate assay (100µl/well). Plates were spun (2 minutes, 1000rpm), supernatant was removed then 100µl glutaraldehyde solution (0.375% in cold PBS) was added for 10 minutes. Finally, plates were spun (2 minutes, 1000rpm) and washed with PBS
- 2- Anti-D antibodies were added and incubated for 1h at room temperature and the
- 10 plate were washed with a buffer solution.
- 3- Recombinant soluble Fc receptors were then added. After an 1h incubation period, the plates were washed. The soluble receptors, FcRIIB (CD32) and FcRIIA (CD16) were produced as a fusion protein by coupling to the GST. The recombinant receptors were purified by affinity chromatography on glutathion agarose.
- 15 PUC119-derived vector containing the FcgRIIA2 cDNA and pcDNA3 vector containing cDNA encoding the FcgRIIIB (NA2 allele) ectodomains were used to construct cDNA encoding recombinant GST fused to the extracellular (EC) region of FcgRIIA (GST-ECIIA) or FcgRIIIB (GST-ECIIB). A two step PCR was performed for generating the GST-ECIIA construct. The following amplimers were used for this
- 20 step : 5'-GCAGCTCCCCCAAAGGCTGTG-3' (sense) and 5'-TTGGACAGTGATGGTCACAGG-3' (antisense). The second step allowed the addition of restriction sites to the PCR product for cloning into the pGEX-2T expression vector (Amersham-Pharmacia Biotech, England). For cloning FcgammaRIIIB EC region, one step PCR was performed using sense (5'-
- 25 TGGATGAATTCCTATTAAGTGATGGTGATGTT-3') and antisense (5'-ATCGGATCCCGACTGAAGATCTC-3') amplimers containing EcoRI and BamHI restriction sites that made possible the cloning of the cDNA into pGEX-2T vector. Fusion proteins (GST-ECIIA, GST-ECIIB) were purified by affinity chromatography onto glutathione agarose beads according to the manufacturer's
- 30 instructions.

4- The binding of the soluble receptor is assessed by adding an antibody labeled with biotine or alkaline phosphatase and specific of the studied receptor. This antibody did not inhibit the binding of the Fc to his receptor. A second anti-IgG and labeled monoclonal antibody is added and the OD are registered with a spectrophotometer.

CLAIMS

1. A method for the preparation of human, humanized or chimæric antibodies or polypeptides comprising Fc region of human IgG with enhanced therapeutic properties and decreased side effects, wherein said method comprises the step consisting of :

 - a) providing candidate human, humanized or chimæric antibodies or polypeptides comprising Fc region of human IgG produced from different cell lines selected from hybridoma, heterohybridoma, animal cell lines or eukaryote microorganisms, transfected with a vector comprising the coding sequence for said antibody or polypeptide,
 - b) testing the binding of said antibodies or polypeptides on Fcgamma receptors including CD16 (FcgammaRIII) and CD32 (FcgammaRIIA and B),
 - c) selecting antibodies or polypeptides which :
 - i) bind specifically or preferentially to a first Fcgamma receptor,
 - ii) bind to both said first Fcgamma receptor and a second Fcgamma receptor,
 - iii) do not bind or bind only weakly to both said first Fcgamma receptor and said second Fcgamma receptor, or
 - iv) bind specifically or preferentially to said second Fcgamma receptor.
2. A method according to claim 1, wherein step c) consists of selecting antibodies or polypeptides which:

 - i) bind specifically or preferentially to CD16,
 - ii) bind to both CD16 and CD32,
 - iii) do not bind or bind only weakly to both CD16 and CD32, or
 - iv) bind specifically or preferentially to CD32.
3. A method according to claim 1 or 2 which further comprise testing the binding to a third or fourth Fcgamma receptor.

4. A method according to one of claims 1 to 3, wherein the binding assays is performed using:

- i) indicator cells from cell lines that express different Fc receptors on their cell surface, or
- ii) recombinant Fc receptors comprising FcγRIIIa ectodomains, Fc receptors derived peptides, or
- iii) in an assay reaction mixture comprising a soluble Fc receptors or fragment thereof.

5. A method according to claim 4, wherein one of the Fc receptors is FcγRIIb.

6. A method according to claim 4, wherein one of the Fc receptors is FcγRIII.

7. A method according to one of claims 1 to 6, wherein the subset of antibodies or polypeptides selected in step c) for the feature (iv) or (ii), more particularly the subset of antibodies or polypeptides selected to bind specifically or preferentially CD32 (iv) or both CD16 and CD32 (ii), are further tested and selected for their ability to trigger or not the inhibitory functions through the human type II receptors for the Fc region of IgG (FcγRIIb/CD32).

8. A method according to one of claims 1 to 6, wherein the subset of antibodies or polypeptides selected in step c) for the feature (iv) or (ii), more particularly the subset of antibodies or polypeptides selected to bind specifically or preferentially CD32 (iv) or both CD16 and CD32 (ii), are further tested and selected for their ability to trigger FcγRIII/CD16 leading to improved ADCC, increased production of cytokines such as Interleukin-2 (IL-2) and/or pro-inflammatory molecules such as Tumor Necrosis Factor alpha (TNFα).

9. A method according to claim 7 or 8 wherein said subset of antibodies or polypeptides are selected for their ability to induce ADCC through the FcγRIII and trigger the inhibitory functions through the FcγRIIb/CD32.

10. A method according to one of claims 7 to 9, wherein the functional assays consist of a calcium mobilization assay and/or a cytokine secretion assay.
11. A method according to one of claims 7 to 10, wherein the functional assays further
5 comprise a specific FcγRIII ADCC assay.
12. A method according to claim 11, wherein the specific FcγRIII ADCC assay consists of the addition of each antibody or polypeptide to a distinct reaction mixture comprising target cells, effector cells expressing FcγRIII and polyvalent IgG, and the
10 determination of target cells lysis percentage.
13. A method according to one of claims 1 to 12, wherein said antibodies or polypeptides are selected as displaying one of the following features:
- 15 i) bind specifically or preferentially to CD16,
ii) bind to both CD16 and CD32,
iii) do not bind or bind only weakly to both CD16 and CD32, or
iv) bind specifically or preferentially to CD32,
are further tested in different culture conditions and different media for improving yield, antibodies or polypeptide properties as featured in i), ii), iii), or iv) and functions.
- 20 14. A method according to one of claims 1 to 13, wherein antibodies or polypeptides comprising Fc region of human IgG are selected for at least one of the following features:
- 25 a) giving a percentage of labeled cells (% positive cells) that represents at least a thirty-fold increase, more preferably at least a forty-fold increase, as compared to the percentage of cells labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD16 (FcγRIIIA) positive indicator cells;
- 30 b) giving a percentage of labeled cells (% positive cells) that represents at least a ten-fold increase, more preferably a twenty-fold increase, as compared to the percentage of cells

labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIB) positive indicator cells;

- 5 c) giving a percentage of labeled cells (% positive cells) that represents at least a five-fold decrease, more preferably a ten-fold decrease, as compared to the percentage of cells labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIA) positive indicator cells;

10

- d) giving a percentage of labeled cells (% positive cells) that represents at least a three-fold decrease, more preferably a five-fold decrease, as compared to the percentage of cells labeled with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIB) positive indicator cells.

15

15. A method according to one of claims 1 to 14, wherein antibodies or polypeptides comprising Fc region of human IgG are selected for at least one of the following features:

- 20 e) giving a mean fluorescence intensity (MFI) value that represents at least a 10-fold increase, more preferably a 15-fold increase, as compared to the MFI value obtained with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIA) positive indicator cells;

25

- f) giving a mean fluorescence intensity (MFI) value that represents at least a 2-fold increase as compared to the MFI value obtained with equivalent doses of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), in an immunofluorescence assay that measures the binding of antibody to CD32 (FcγRIIB) positive indicator cells;

30

g) a mean fluorescence intensity (MFI) value that represents at least a five-fold decrease, as compared to the MFI value obtained when equivalent doses of anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), is tested in an immunofluorescence assay that measures the binding of antibody to CD16 positive indicator cells;

5

h) a mean fluorescence intensity (MFI) value that represents at least a five-fold decrease as compared to the MFI value obtained when the same dose of the anti-CD20 monoclonal antibody, Rituxan (Rituximab) (clinical grade), is tested in an immunofluorescence assay that measures the binding to CD32 (FcgammaRIIB) positive indicator cells.

10

16. A method according to one of claims 1 to 15, wherein antibodies or polypeptides comprising Fc region of human IgG are selected for at least one of the following features:

15 - an elevated mean fluorescence intensity (MFI) and a strong percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD16 (FcgammaRIII) positive indicator cells.

20 - a low mean fluorescence intensity (MFI) and a low percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD16 (FcgammaRIII) positive indicator cells.

25

- an elevated mean fluorescence intensity (MFI) and a strong percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD32 (FcgammaRIIB) positive indicator cells.

30 - a low mean fluorescence intensity (MFI) and a low percentage of labeled cells (% positive cells) in an immunofluorescence assay that measures the binding to CD32 (FcgammaRIIB) positive indicator cells.

17. A method of purification of antibodies exhibiting binding properties to FcgammaRI and FcgammaRII such as those defined in one of claims 14 to 16 comprising a step consisting of enriching said antibodies by using immobilized soluble forms of FcgammaRI termed FcgammaR-immunoadsorbents, comprising the extracellular regions of the receptors.

5

18. A method according to claim 17 to enrich for antibodies that efficiently bind FcgammaRIII and/or FcgammaRII, wherein said method comprises the retention of antibodies on said FcgammaR-immunoadsorbents, and the elution of preparation containing "high" FcgammaR binders antibodies.

10

19. A method according to claim 17 to enrich for antibodies that poorly bind FcgammaRI and/or FcgammaRII or that do not bind FcgammaRIII and/or FcgammaRII, wherein said method comprises the collection of the effluent fraction of said FcgammaR-immunoadsorbents, allowing the obtention of preparations containing "low" FcgammaR binders antibodies.

15

20. A composition comprising at least 80%, preferably at least 95%, of antibodies obtainable by the method according to one of claims 1 to 20.

20

21. A composition comprising at least 80%, preferably at least 95% of antibodies or peptides displaying at least one (for example 2) of the following features:

a) a MFI higher than 500, more preferably 700, and a percentage of labeled cells higher than 90 %, more preferably 95 %, in an immunofluorescence assay that measures the binding of 5, 10, 25 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD16 (FcgammaRIII) positive indicator cells (characterized by a number of FcgammaRIII receptors lower than 10^5 /cell); An example of this is the R297 antibody. Another example is the 4B12 antibody.

25

b) a MFI lower than 80, more preferably 20, and a percentage of labeled cells lower than 5 % , more preferably 2 % , in an immunofluorescence assay that measures the binding of 5, 10, 25 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD16 (FcgammaRIII) positive indicator cells (characterized by a number of 5 FcgammaRIII receptors lower than 10^5 /cell); An example of this is the AD1 antibody. Another example is the B11 antibody.

c) a MFI higher than 50, more preferably 100, and a percentage of labeled cells higher than 80 % , more preferably 90 % in an immunofluorescence assay that measures the binding of 10, 25, 50 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD32 (FcgammaRIIB) positive indicator cells (characterized by a number of FcgammaRIIB receptors lower than 5×10^4 /cell). An example of this is the R297 antibody. Another example is the 4B12 antibody.

d) a MFI lower than 50, more preferably 20, and a percentage of labeled cells lower than 5 %, more preferably 2 % , in an immunofluorescence assay that measures the binding of 10, 25, 50 micrograms/ml antibody (containing less than 2% aggregates as determined by HPLC) to 5×10^5 CD32 (FcgammaRIIB) positive indicator cells (characterized by a number of FcgammaRIIB receptors lower than 5×10^4 /cell). An example of this is the AD1 20 antibody. Another example is the B11 antibody.

22. A composition comprising at least 80%, preferably at least 95% of antibodies or peptides displaying the feature as defined in c) in claim 21.

25 23. A composition comprising at least 80%, preferably at least 95% of antibodies or peptides displaying the features as defined in b) and c) in claim 21.

24. A composition comprising at least 80%, preferably at least 95% of antibodies or peptides displaying the features as defined in a) and c) in claim 21.

25. A composition comprising at least 80%, preferably at least 95% of antibodies or peptide displaying the feature as defined in d) in claim 21.
26. A composition comprising at least 80%, preferably at least 95% of antibodies or peptide displaying the features as defined in b) and d) in claim 21.
27. A composition comprising at least 80%, preferably at least 95% of antibodies or peptide displaying the features as defined in a) and d) in claim 21.
28. Use of a composition according to one of claims 22 to 24 to manufacture a medicament for treating auto-immune diseases or other antibody-related pathologies such as allergic immunization.
29. Use of a composition according to one of claims 22 to 24 to manufacture a medicament for treating allergies and IgE-dependent Type I hypersensitivity.
30. Use of a composition according to one of claims 22 to 24 to manufacture a medicament for treating allergies.
31. Use according to one of claims 22 to 24 for treating asthma, allergic rhinitis, allergic sinusitis, anaphylactic syndrome, urticaria, angioedema, atopic dermatitis, allergic contact dermatitis and erythema.
32. Use of a composition according to one of claims 25 to 27 to manufacture a medicament for treating cancer.
33. Use of a composition according to one of claims 22 to 24 to manufacture a medicament for the regulation of hematopoietic cell proliferation dependent on Receptor Tyrosine Kinases (RTKs) by FcγRIIB.

34. Use according to claim 33, wherein said antibodies are bifunctional molecules such as anti-RTK IgG antibodies with optimized Fc region capable of efficiently co-aggregating RTKs with FcgammaRIIB, to manufacture a medicament for blocking the proliferation of FcgammaRIIB+ tumor cells.

5

35. Use of a composition according to one of claims 20 to 27 to manufacture a medicament, wherein the antibody recognizes antigens expressed on or bound to the cell surface of target cells.

10

36. Use according to claim 35, wherein said antigens include allo-antigens, transplantation antigens, self-antigens such as class I and class II Major Histocompatibility Antigens (HLA), FcgammaRI, FcgammaRIIA, FcgammaRIII molecules, FcalphaR, FcepsilonRI, B cell receptor, T cell receptor, tumor antigens such as CD20, Her2/NEU, CEA, GD2, allergen such as phospholipase A2, and IgE.

15

37. Use of a composition according to one of claims 20 to 27, wherein the antibodies recognize Rhesus D antigen to manufacture a medicament for treating or preventing Rhesus alloimmunization of Rh negative patients, leading to the Hemolytic Disease of the New Born (HDNB).

20

38. Use of a composition according to one of claims 20 to 27, wherein the antibodies recognize Rhesus D antigen to manufacture a medicament for treating or preventing Idiopathic Thrombocytopenic Purpura (ITP) .

25

39. Use of a composition according to one of claims 20 to 27, wherein the antibodies HLA Class II molecules to manufacture a medicament for treating or preventing cancers, autoimmune diseases, or graft rejections.

30

ABSTRACT

- 5 The present invention relates to a method for the production and the selection of human or chimæric or humanized antibodies or molecules that comprise the Fc region of human IgG, capable of modulating the activity of one or several particular Fc receptors, such as the triggering of inhibitory functions through the human type II receptors of IgG (FcγRII/CD32).

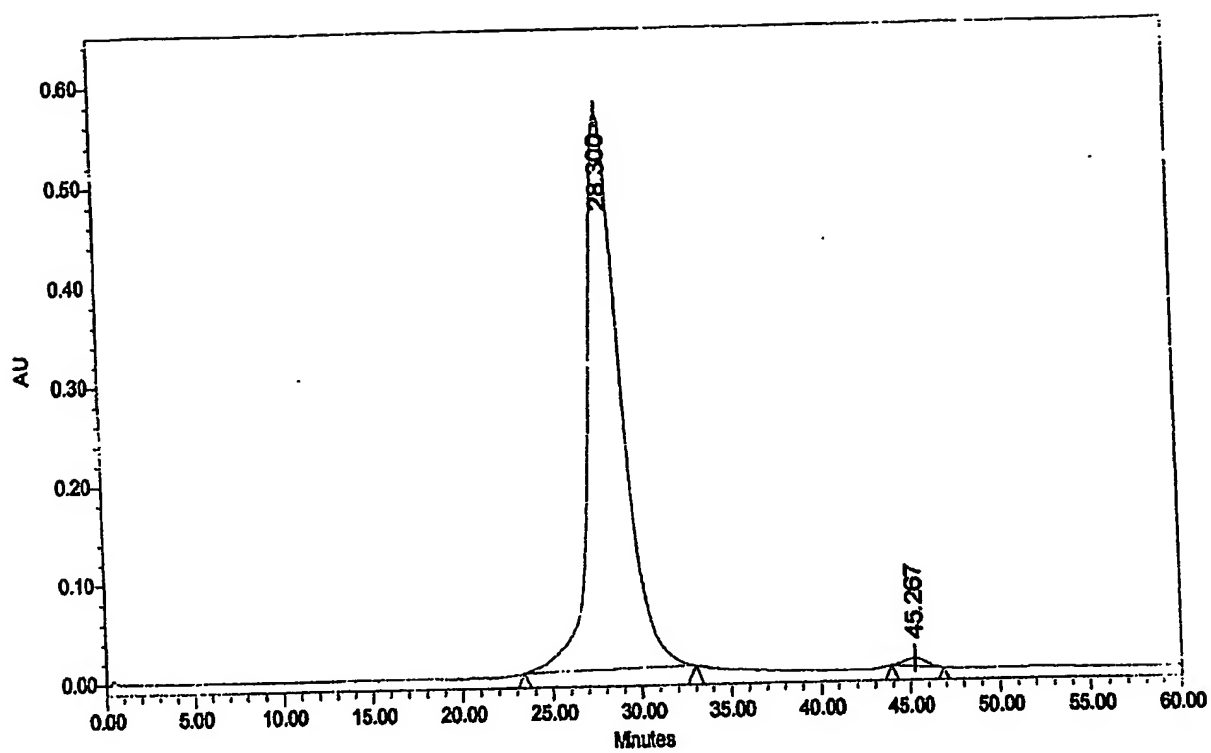


FIGURE 1

Oligosaccharide composition of different monoclonal antibodies determined by HPCE-LIF

Structures	Anti-D		Anti-HLA-DR	
	R297, YB2/0	AD1, Heterohybrid	4B12, YB2/0	B11, CHO
Fucosylated	22	67		
Sialylated	0	30		
A2F	ND	8,7		
A2FB	ND	ND		
A1F	ND	21,3		
A1FB	ND	ND		
G2FB	ND	ND		
G2F	5,7	16,8		
G2	8,3	2		
G1F	20,4	18,2		
G1	35,7	1,7		
G0F	4,8	1,7		
G0	27,8	0,5		

ND: not detected

Different forms of glycans :

A: sialilated

G0 : agalactosylated

G1 : monogalactosylated

G2 : bigalactosylated

F : fucosylated

B : bisecting GlNAc

FIGURE 2

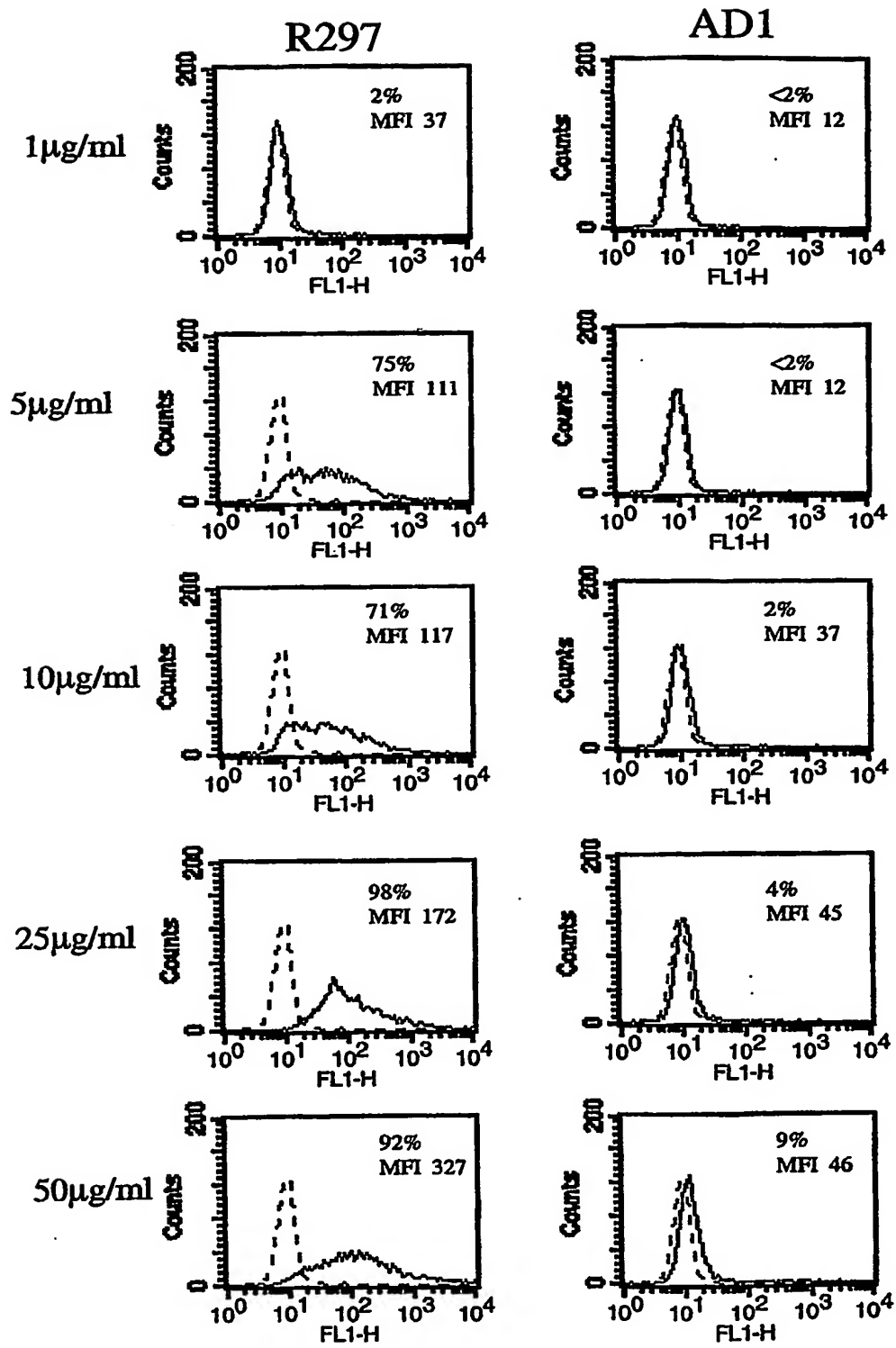


FIGURE 3

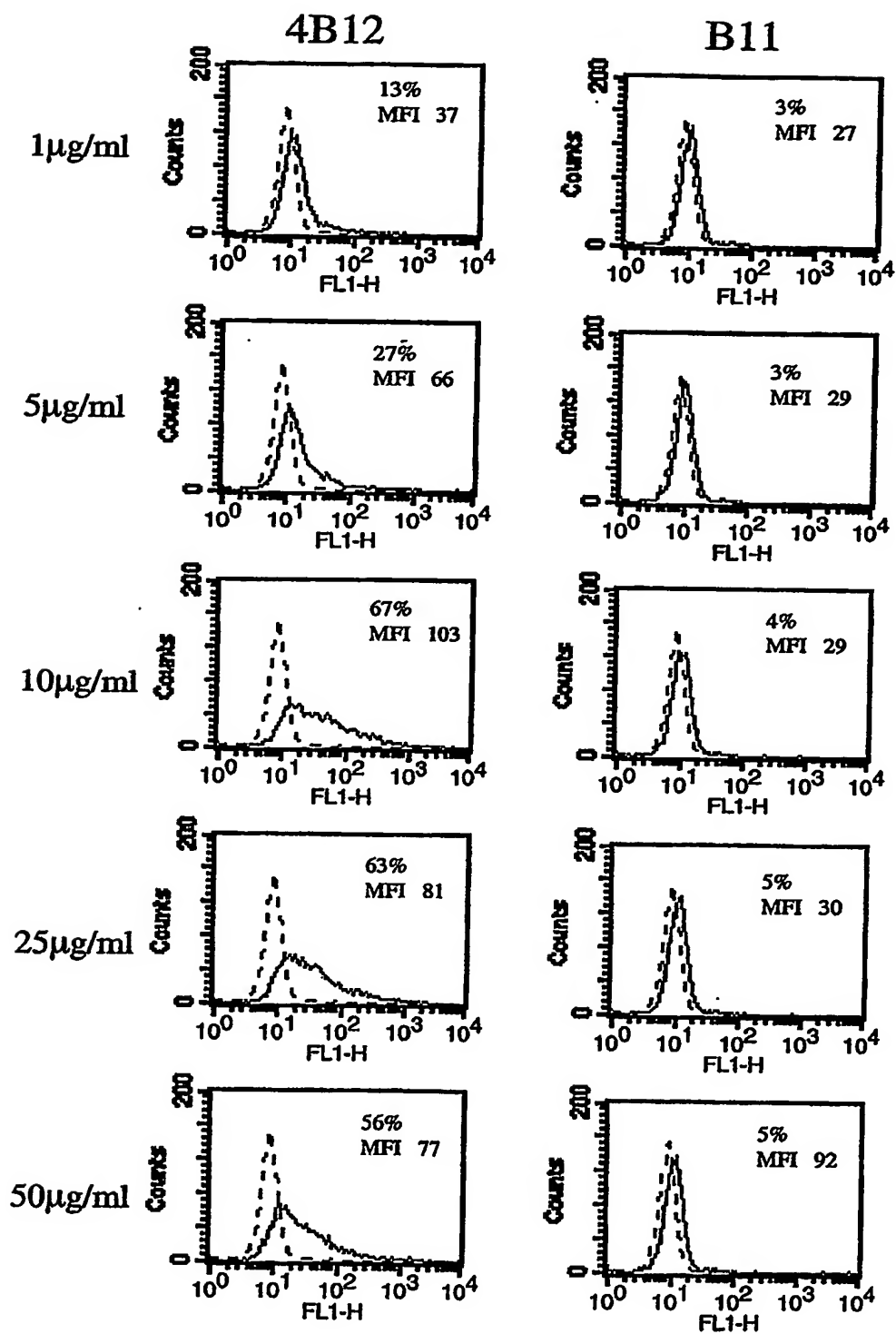


FIGURE 4

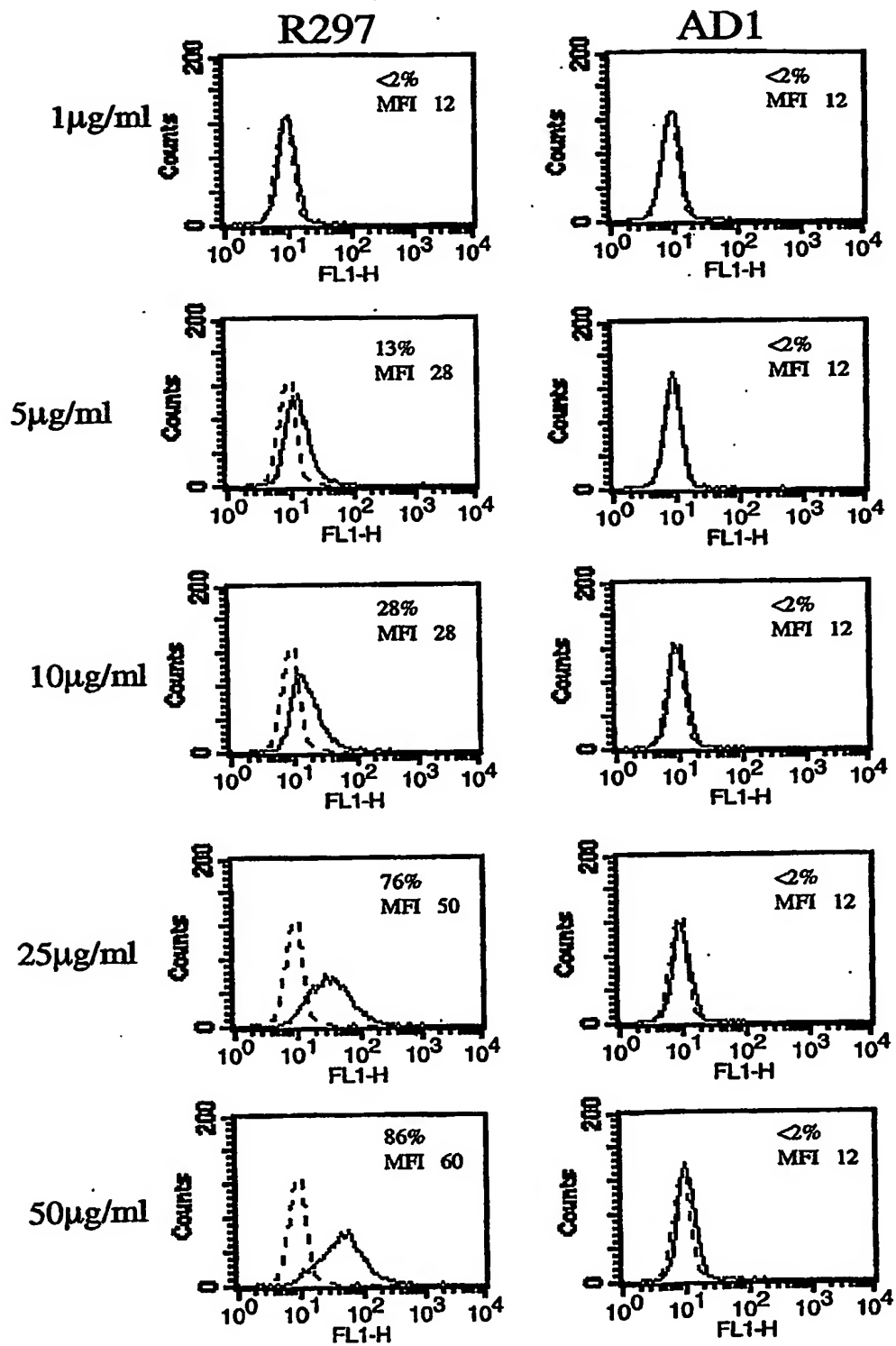


FIGURE 5

6 / 13

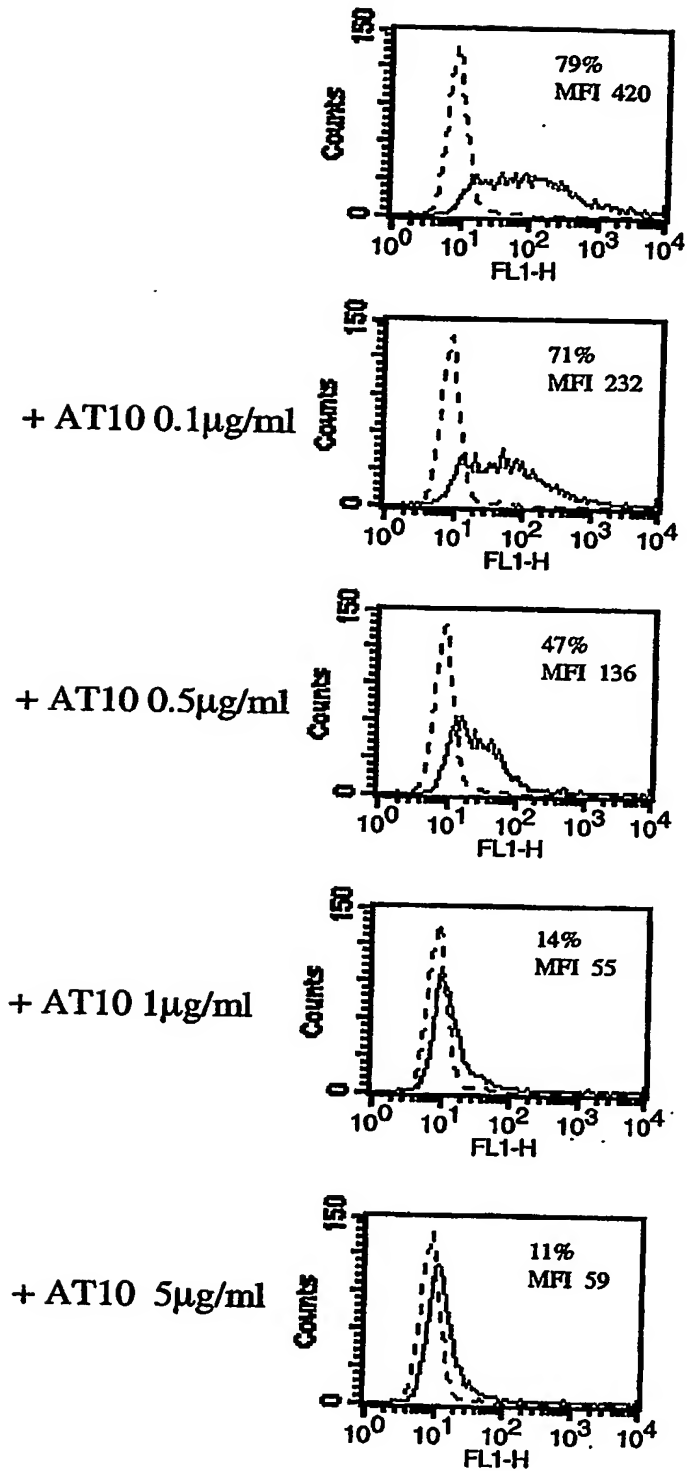


FIGURE 6

7 / 13

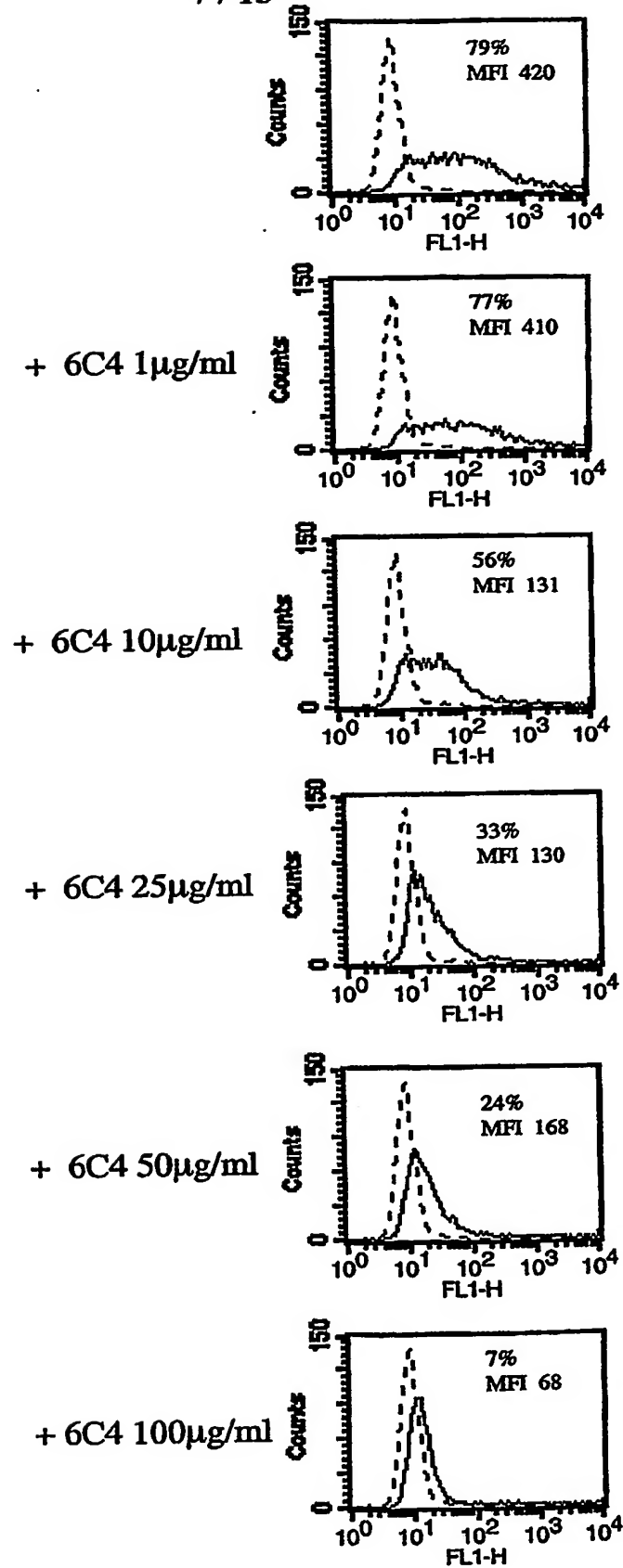


FIGURE 7

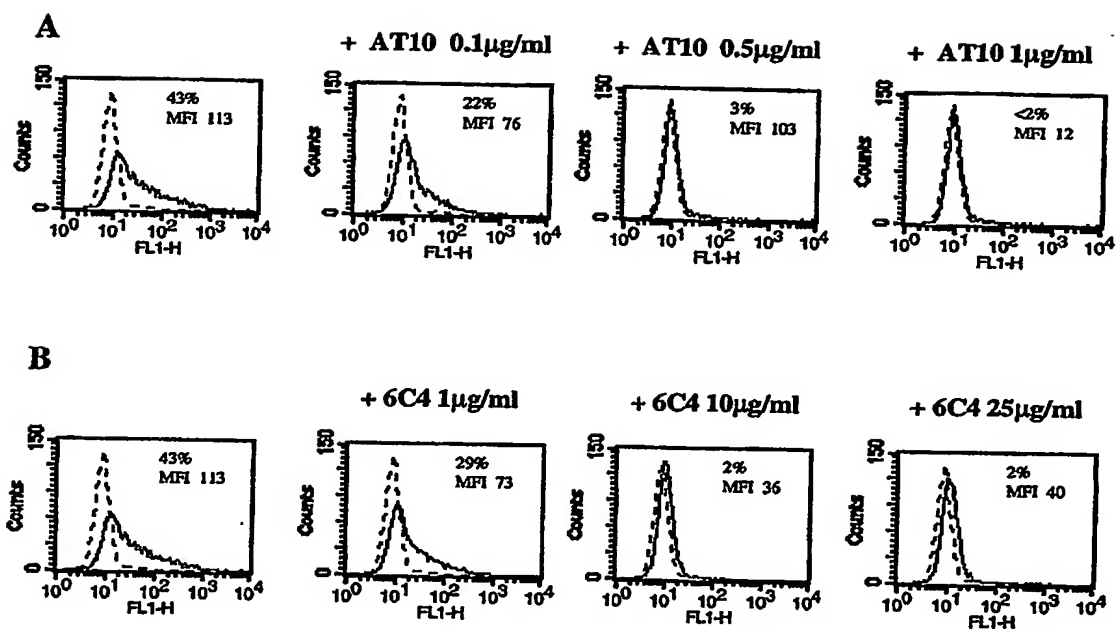


FIGURE 8

9 / 13

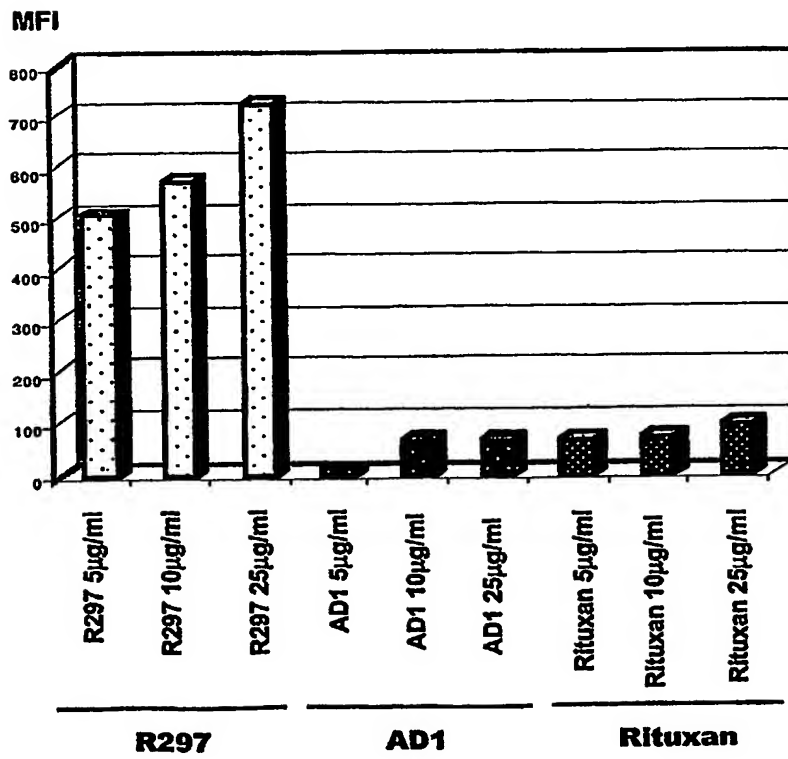
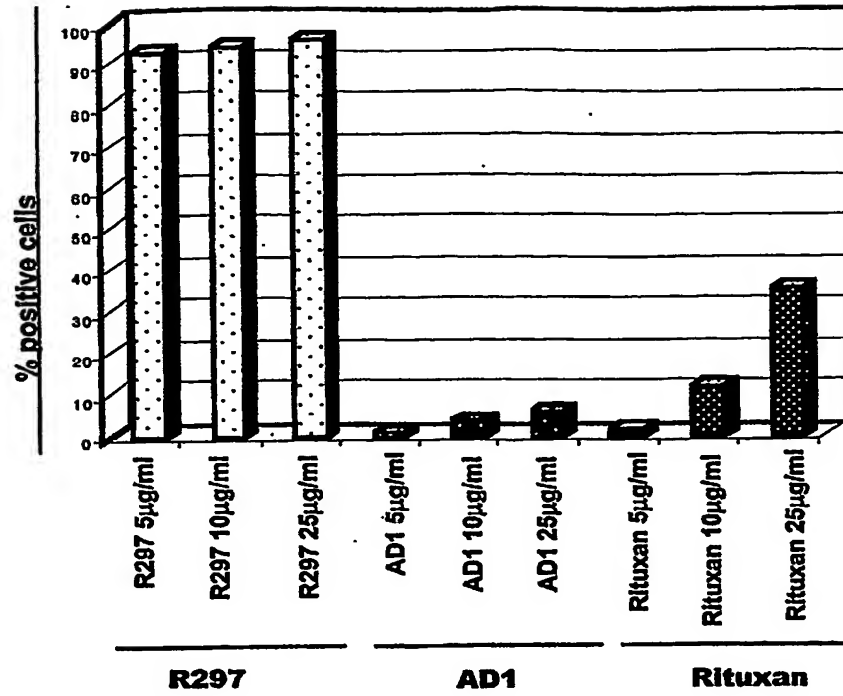
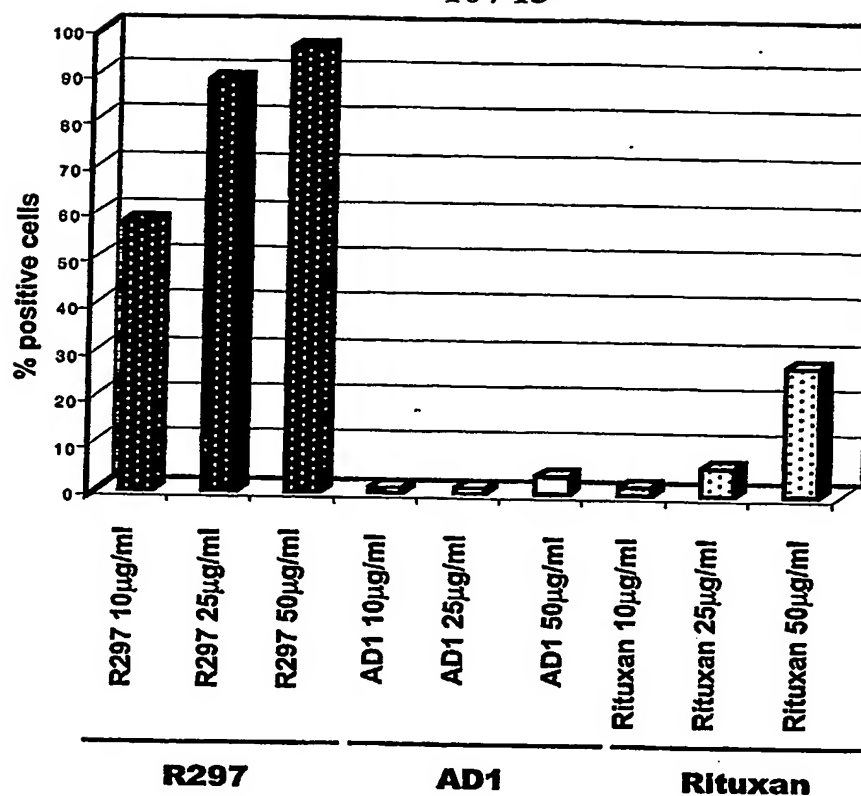


FIGURE 9

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MFI

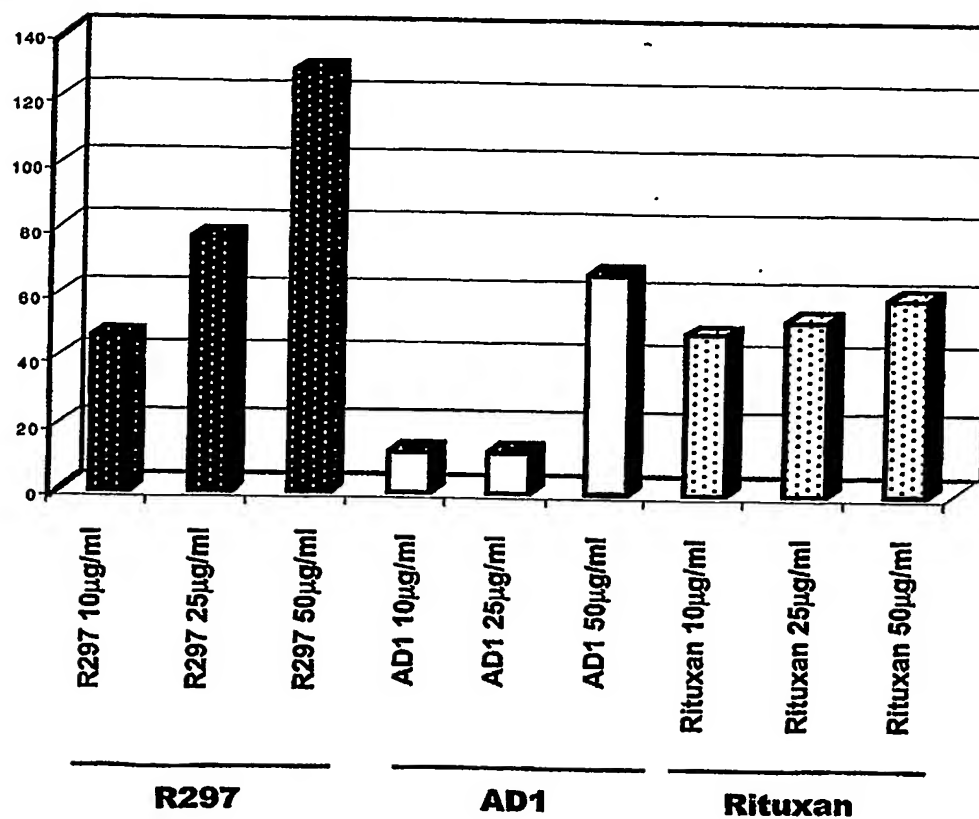


FIGURE 10

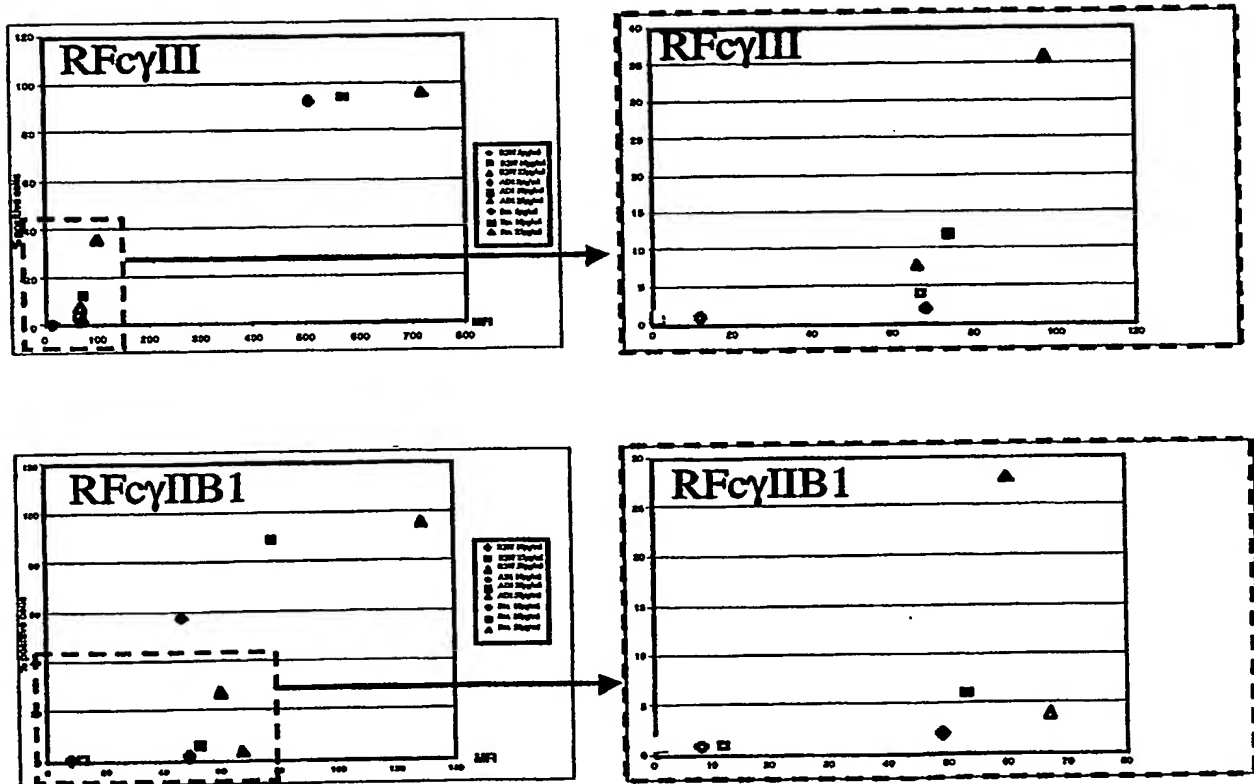


FIGURE 11

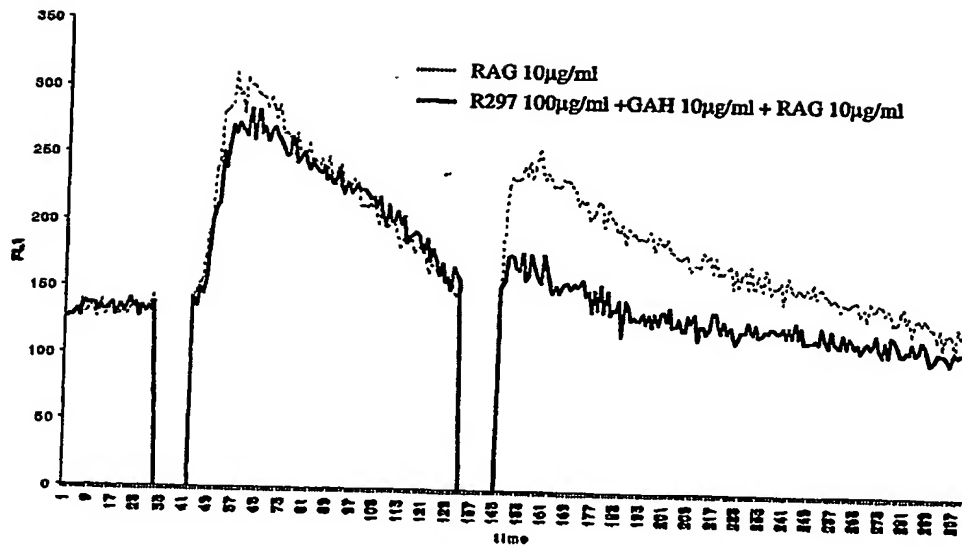


FIGURE 12

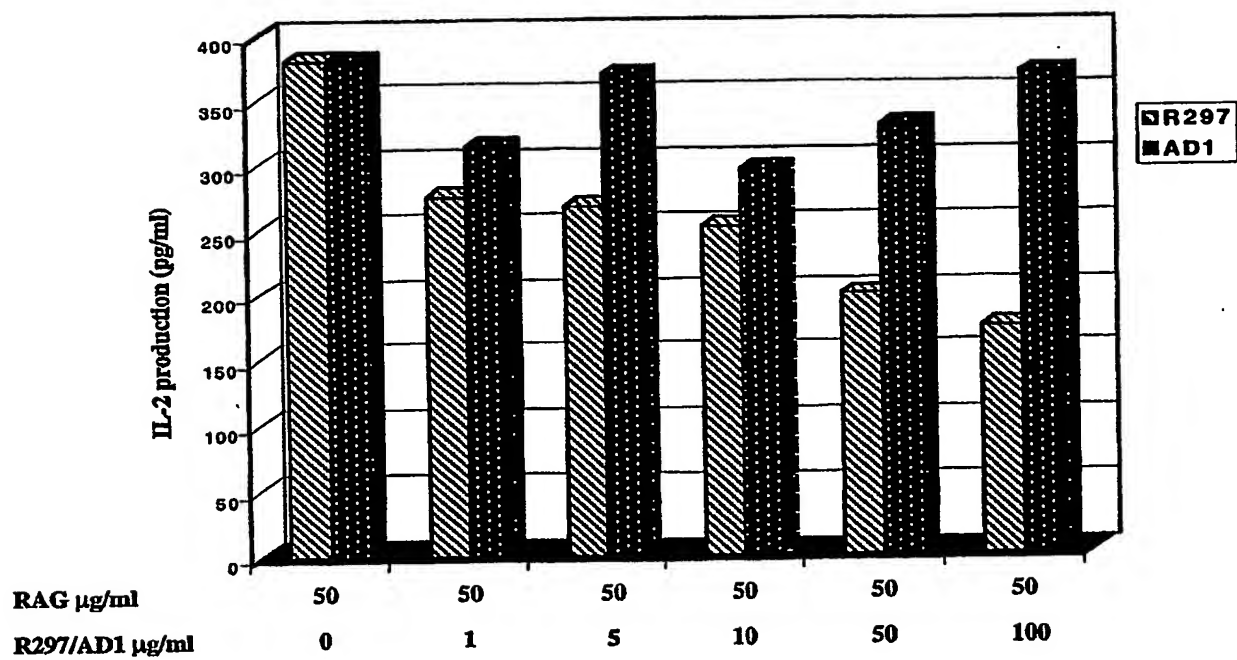


FIGURE 13

SEQUENCE LISTING

<110> LABORATOIRE FRANCAIS DU FRACTIONNEMENT ET DES
BIOTECHNOLOGIES (LFB)

<110> INSTITUT NATIONAL DE LA SANTE ET DE LA RECHERCHE MEDICALE
(INSERM)

<120> THERAPEUTIC PRODUCTS WITH ENHANCED ABILITY TO IMMUNOMODULATE CELL
FUNCTIONS

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<170> PatentIn version 3.2

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PCT/IB2004/001390

